

Annual Review of Entomology

Non-Bee Insects as Visitors and Pollinators of Crops: Biology, Ecology, and Management

R. Rader,^{1,*} S.A. Cunningham,² B.G. Howlett,³
and D.W. Inouye^{4,5}

¹School of Environmental and Rural Science, University of New England, Armidale, New South Wales 2351, Australia; email: rrader@une.edu.au

²Fenner School of Environment and Society, College of Science, The Australian National University, Canberra ACT 2601, Australia

³The New Zealand Institute for Plant and Food Research Limited, Christchurch 8140, New Zealand

⁴Department of Biology, University of Maryland, College Park, Maryland 20742, USA

⁵Rocky Mountain Biological Laboratory, Crested Butte, Colorado 81224, USA

Annu. Rev. Entomol. 2020. 65:391–407

First published as a Review in Advance on
October 14, 2019

The *Annual Review of Entomology* is online at
ento.annualreviews.org

<https://doi.org/10.1146/annurev-ento-011019-025055>

Copyright © 2020 by Annual Reviews.
All rights reserved

*Corresponding author

Keywords

Diptera, Coleoptera, Lepidoptera, flower-visitor, crop pollination, pollinator efficiency

Abstract

Insects other than bees (i.e., non-bees) have been acknowledged as important crop pollinators, but our understanding of which crop plants they visit and how effective they are as crop pollinators is limited. To compare visitation and efficiency of crop-pollinating bees and non-bees at a global scale, we review the literature published from 1950 to 2018 concerning the visitors and pollinators of 105 global food crops that are known to benefit from animal pollinators. Of the 105 animal-pollinated crops, a significant proportion are visited by both bee and non-bee taxa ($n = 82$; 77%), with a total gross domestic product (GDP) value of US\$780.8 billion. For crops with a narrower range of visitors, those that favor non-bees ($n = 8$) have a value of US\$1.2 billion, compared to those that favor bees ($n = 15$), with a value of US\$19.0 billion. Limited pollinator efficiency data were available for one or more taxa in only half of the crops ($n = 61$; 58%). Among the non-bees, some families were recorded visiting a wide range of crops (>12), including six families of flies (Syrphidae, Calliphoridae, Muscidae, Sarcophagidae,

ANNUAL REVIEWS CONNECT

www.annualreviews.org

- Download figures
- Navigate cited references
- Keyword search
- Explore related articles
- Share via email or social media

Tachinidae, and Bombyliidae), two beetle families (Coccinellidae and Nitidulidae), ants (Formicidae), wasps (Vespidae), and four families of moths and butterflies (Hesperiidae, Lycaenidae, Nymphalidae, and Pieridae). Among the non-bees, taxa within the dipteran families Syrphidae and Calliphoridae were the most common visitors to the most crops, but this may be an artifact of the limited data available. The diversity of species and life histories in these groups of lesser-known pollinators indicates that diet, larval requirements, and other reproductive needs will require alternative habitat management practices to bees.

INTRODUCTION

Most of the crops grown around the world benefit from insect pollination either for the direct production of commodities (i.e., fruit or seed consumed by humans) or in the production of seed required to cultivate the vegetative parts of the crop for consumption (i.e., vegetable seed). The 105 most widely planted crops globally that benefit from insect pollination have greater than US\$800 billion gross economic value and include many of the most popular fruit, vegetable, and nut commodities consumed worldwide, such as melons, avocados, berries, and almonds (63; <http://www.fao.org/faostat/en/#home>).

Both wild and managed pollinators provide significant crop pollination services and often complement each other in commercial production systems (8, 32). Bees are the most commonly studied crop pollinators and are thought to dominate many crop pollinator assemblages, in terms of both abundance and diversity (32, 61, 111). Several bee species have been introduced to provide crop pollination services, some of which are managed. These managed species include the honey bees (Apini); stingless bees (Meliponini); *Bombus* spp.; and a few solitary bees, including Alfalfa leaf cutter bees (*Megachile rotundata*), *Osmia* bees (*Osmia* spp.), and Alkali bees (*Nomia melanderi*). The wild, unmanaged bees that provide crop pollination services include over 700 different species, both social and solitary (32, 61).

There are numerous wild pollinators that are not bees. Myriad non-bees constitute the diverse communities of invertebrate and vertebrate taxa that visit wild plants (78) and cultivated crops (84). Like bees, several non-bee taxa have also been introduced outside of their native host ranges to facilitate the commercial production of specialist crops, such as the *Elaeidobius kamerunicus* weevil for oil palm and fig wasps for fig fruits (11, 107). In addition, some fly taxa are now being specifically reared commercially and managed for the purpose of crop pollination (<http://www.polyfly.es>) and rearing protocols are available for some species, such as Syrphid flies (77). While wild non-bee taxa were recorded in a 2007 synthesis (63), this study did not explore the diversity of non-bee taxa that pollinate globally important food crops, and many studies have since added to our understanding of these lesser-known pollinators.

Different pollinators vary significantly in their pollination performance across crops (32, 84). Understanding which taxa are most effective at transferring pollen is important, as these are the most likely contributors to pollinator-dependent crop yields. While most of these crops have open, easily accessible flowers that are visited by generalist pollinator taxa, some major crop groups have specialized flower morphologies that are likely to limit the effectiveness of some pollinators. For example, poricidal anthers (e.g., many Solanaceae, including tomato and eggplant) require vibration to release pollen, and flower-tripping mechanisms (e.g., papilionoid legumes) are thought to be best pollinated by bee visitors (15). Other crops are pollinated by non-bee insects including, some for which the major pollinator has complex breeding site requirements, such as oil palm (*Elaeis guineensis*), figs (*Ficus carica*), and jackfruit (*Artocarpus heterophylla*) (31, 103),

or habitat requirements, such as fruit husks or decomposing leaf litter to optimize pollination in cocoa (*Theobroma cacao*) (25). Others, such as durian, have flowers that dehisce at night, when bees are generally not in flight (68).

At present, it is challenging to draw general patterns regarding crop pollinator assemblages and their efficiency for all but a small number of crops because efficiency data are lacking. With increasing human population growth and global food demands, understanding the factors that influence the yields of the world's food crops is critical to sustaining ongoing production. In this article, we review the literature to compare and contrast the contribution by bees and non-bees to global food crops. We evaluate the identity and richness of floral visitors, their effectiveness as crop pollinators, and the opportunities and challenges that will likely influence their management in agricultural environments.

We ask the following questions:

1. How diverse are the assemblages of non-bee flower visitors that visit the 105 major animal pollinated crops, and how does this compare to bees?
2. What data are available on the importance of non-bee insects as pollinators?
3. What is the economic value of bee- and non-bee-visited crops to the global economy?

LITERATURE SEARCH AND ANALYSES

Methods

We conducted a literature search based on the supplementary information in the work of Klein et al. (63), selecting the 105 world crops (**Supplemental Table 1**) that benefit from animal pollination for direct production of commodities (i.e., fruit or seed consumed by humans) or that indirectly benefit through the increased production of seed required to cultivate the vegetative parts of the crop for consumption (i.e., vegetable seed). Crops for which the consumed parts are primarily vegetatively propagated were excluded (e.g., potatoes). To minimize duplicate handling of search results for congeneric crops, we grouped crop queries by genera. We grouped together common names and included generic synonyms published in Reference 63 to achieve an exhaustive search and removed ubiquitous words from the search string to improve search precision. The target crops (**Supplemental Table 1**), crop search terms, and associated search notes are listed in **Supplemental Table 2**. For each crop group, we searched the Scopus database from 1950–November 1, 2018 using the query (TITLE-ABS-KEY(pollinat* AND ('crop common names' OR 'crop genus'))). In total, our search queries retrieved 12,680 results. The qualitative results of our flower-visitor family census were unaffected by duplicated references and linked articles. We retained Klein et al.'s list of taxa in our study, which includes additional crop-visitor taxa references not revealed in our search.

Search results were filtered at three levels: (a) title and journal, (b) abstract, and (c) full text. Search results containing unfamiliar specific epithets for crop genera were checked against The Plant List (<http://www.theplantlist.org>) to determine if names were synonyms of target crops. We included visitors to cultivated and naturalized crops as well as crops grown as sentinel or phytometer plants. Non-English titles, abstracts, methods, and results were translated using Google Translate (<https://translate.google.com.au>). Articles were excluded if the titles indicated that the material was not relevant; the majority of these articles focused on nontarget congeneric plant taxa or taxa with shared common names, or were molecular studies published in molecular biology and related discipline journals. For the remainder of the search results, abstracts were scanned for relevance to pollination of the targeted crops. Articles that were clearly focused on animal pollination or were ambiguous in regard to animal pollination were viewed as full text.

Supplemental Material >

We examined the methods to determine if flower visitors were observed directly, or if indiscriminate methods such as pan-trapping or extensive sweep-netting were used to populate crop-visitor lists, in which case such studies were excluded. If articles explicitly stated that particular flower-visiting taxa did not contact the reproductive structures of the flowers, then we did not include these taxa. Some studies did not clearly define their usage of phrases such as flower visitors, but we included these taxa nonetheless. In total, we used data from 1,022 publications in this study.

We used the Global Biodiversity Information Facility (GBIF) species match application program interface (API) (<https://www.gbif.org>) to retrieve taxonomic information for flower-visiting taxa on February 20, 2019. We manually evaluated all returned nonexact, fuzzy logic-based matches against the taxon name recorded from the search. For names not in the GBIF, we checked the source article for taxonomic information, which, if unpublished, was obtained manually through GBIF genera search queries or Google Scholar.

Crop production values were obtained from the Food and Agriculture Organization of the United Nations (FAO) agricultural database (<http://www.fao.org/faostat/en/#home>) for the year 2016 (**Supplementary Table 7**). As the FAO lists multiple commodities derived from single crops, we selected only the single most relevant commodity to represent each of our crop groups. As the contributions of individual crops were unknown from some crop groupings listed in the FAOSTAT database, many crops were grouped as not elsewhere classified (NES) and were excluded in calculations of gross domestic product (GDP) economic value; therefore, estimates of value should be treated as conservative.

Supplemental Material >

Results

Crop flower visitor diversity. At least 20 different animal orders visited the flowers of the 105 listed crops (**Figure 1**; **Supplementary Table 3**). Although insects were the most commonly observed, visitors included vertebrates, such as birds and bats, and non-insect arthropods, such as spiders. Of the 105 crops that benefit from animal pollination, most of the crops are visited by both bee and non-bee taxa ($n = 82$; 77%). Hymenopterans, most of which were bees (**Figure 1**), visited the widest range of crops overall (93%).

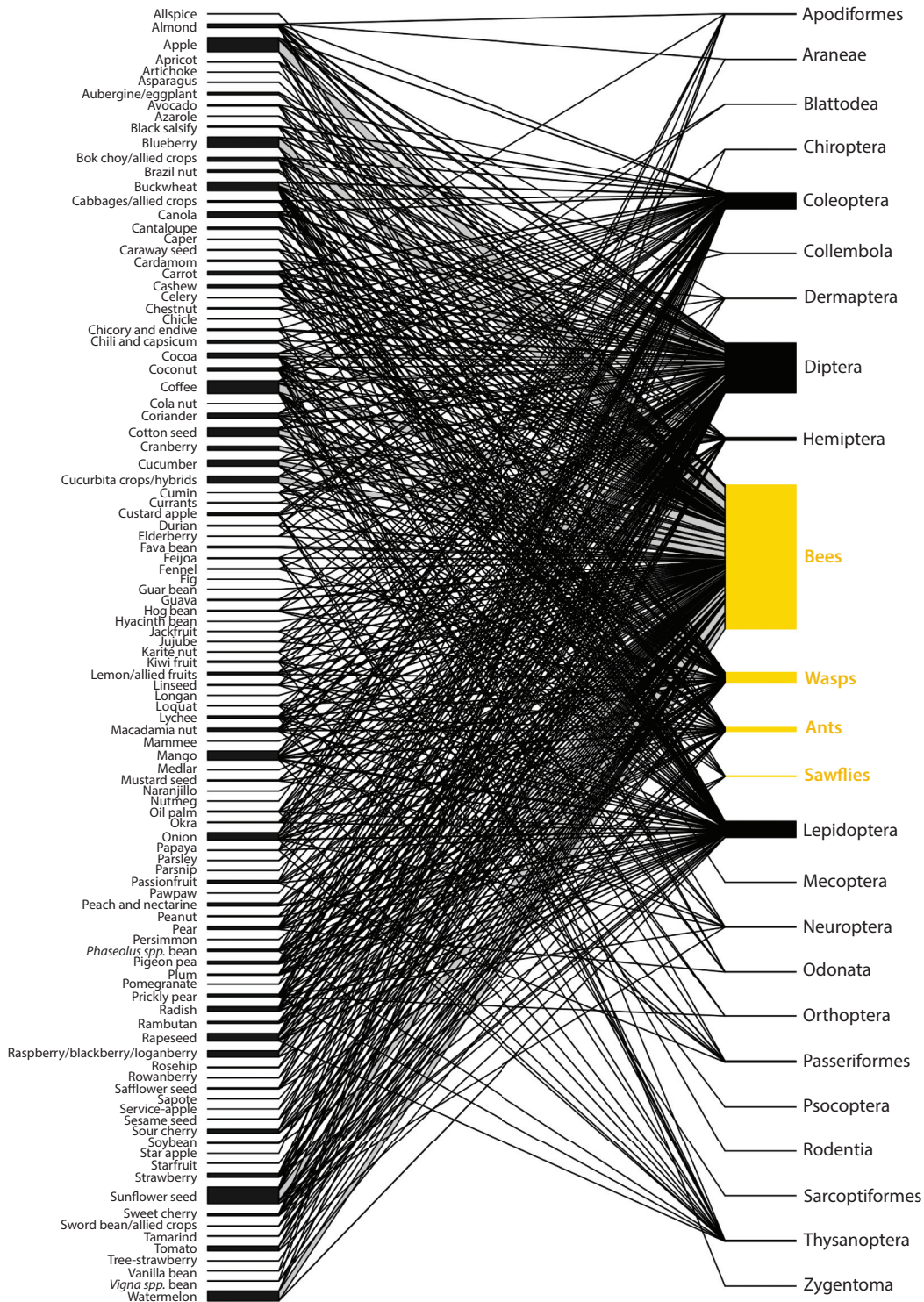
Among the non-bee hymenopterans, Formicidae (ants) were found to visit a large proportion of crops (30%).

Diptera was the second most frequent flower-visiting order (72% of crops), followed by Lepidoptera (54%) and Coleoptera (51%). The fly family Syrphidae (hoverflies) was the most frequent non-bee family, visiting over half of the crop species, followed by the blow flies, in the family Caliphoridae (**Figure 2**).

Coffee was visited by the highest number of insect orders ($n = 10$), followed by buckwheat, bok choy, and allied crops (e.g., Chinese cabbage) and coconut ($n = 9$). Onion and coriander were visited by eight animal orders. Mango, sunflower, cocoa, and coffee had the greatest diversity of families, ranging between 45 and 59 different animal families visiting each of these crops (**Supplementary Tables 3–5**).

While most crops were visited by both taxa, our review identified a small number of major crop species that were visited solely by bees ($n = 15$) or by non-bees ($n = 8$). The crops visited by both bees and non-bees have a total GDP value of US\$780.8 billion. The crops visited solely by bees are valued at US\$19.0 billion, and those visited solely by non-bees at US\$1.2 billion (<http://www.fao.org/faostat/en/#home>; **Supplementary Tables 6 and 7**).

Pollinator effectiveness. Compared to visitation studies, we found relatively few studies that tested differences in pollinator effectiveness among flower visitors (**Table 1**). Efficiency was



(Caption appears on following page)

Figure 1 (Figure appears on preceding page)

A crop–flower visitor network. The width of the bars is based on the number of taxa found to visit that crop group in the literature reviewed; the wider is the bar, the more taxa visited that crop. The yellow bars and labels represent the groups that make up Hymenoptera.

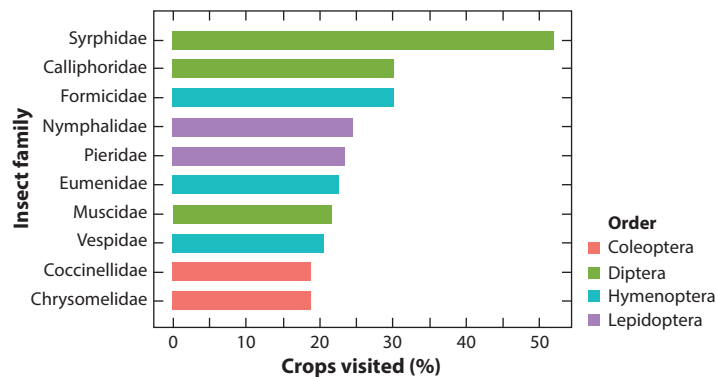


Figure 2

Top 10 non-bee flower visitors by percentage of major crops visited.

measured as pollen deposition on floral stigmas (36 crops for bees, 19 for non-bees), fruit set after caged pollinator or enclosure trials (39 crops for bees, 15 for non-bees), estimates of pollen carried in bee corbiculae or nests (23 crops for bees only), or pollen carried on visitor bodies (not including scopa or corbiculae) (36 crops for bees, 24 crops for non-bees). At least one of the metrics for measuring non-bee effectiveness was only measured in 30 crops. The two effectiveness metrics that included direct estimates of pollen delivered to plants (i.e., fruit set and pollen deposition of stigmas), as opposed to indirect measures of pollen transfer (i.e., body pollen, pollen carried in scopa/corbiculae), were estimated in only 15 crops for non-bees.

Discussion

Our review of the literature indicates that among the many and diverse non-bee pollinators, flies (especially from the families Syrphidae and Calliphoridae; see Table 2) are the most common and visit the widest range of crops. As flies and bees belong to entirely different orders, there are

Table 1 Number of crops for which data are available on effectiveness of non-bee flower visitors as pollinators

Method of estimating effectiveness or importance	Number of crops for which data are available	
	Bees	Non-bees
Single-visit (or multiple-visit) pollen deposition	36	19
Seed or fruit set in cage trials	39	15
Body pollen	36	24
Corbicula and scopa or nest pollen (bees only)	23	NA
Other methods (e.g., correlations of open fruit set with visit rate)	17	9

Abbreviation: NA, not applicable.

Table 2 Life history traits of five fly species that commonly visit a variety of crops

Species	Number of crops visited	Larval life history	Life cycle, generations, and adult longevity	Adult seasonality	Hibernation	Habitat and geographical range	Fecundity	Migratory
<i>Eristalis tenax</i> (Linnaeus, 1758)	28	Inhabit broad range of aquatic and semi-aquatic habitats, e.g., manure, compost, slurry	Typically multiple generations (>2/year) (23) Adult female lifespan is 3 months in the lab without hibernation Egg to adult in 24–36 days (at approximately 21.5°C) (77)	Almost entire year, including mild winter days	As adults	All continents except Antarctica, common in intensive agro-ecosystems (98)	3,000 eggs/female in 2 months (77)	Highly
<i>Episyrphus balteatus</i> (De Geer, 1776)	24	Feed on many aphid species, as well as Coccoidea, Phyllaxeridae, and Psyllidae (36) Found on a wide range of plant species	Typically multiple generations (>2/year) in Europe (23) Egg to adult in 21.2 days (at 26.6°C) (46) Adult female longevity of 38 days or more (greenhouse) (35)	In Europe, ~10 months/year Adults active on milder winter days	As adults	Occurs across a wide range of habitats, including agro-ecosystems (96); present across Europe, North Africa, Asia, and Australia	780 eggs/female in greenhouse (35)	Highly

(Continued)

Table 2 (Continued)

Species	Number of crops visited	Larval life history	Life cycle, generations, and adult longevity	Adult seasonality	Hibernation	Habitat and geographical range	Fecundity	Migratory
<i>Eristalinus aeneus</i> (Scopoli, 1763)	10	Inhabit various aquatic habitats (e.g., ponds, coastal lagoons, rivers, irrigation ditches), including hypersaline environments, dung, and sewage	Typically multiple generations (>2/year) in Europe (23) Female average lifespan is 31.1 days in the lab Egg to emergence is 22.5 days at 20–22°C (76)	Active approximately 5–6 months during warmer months in Europe	As adults	Europe, Africa, Asia, Australia, and North Africa Found in a wide variety of habitats but restricted to coast in northern Europe	Not available	Non (23)
<i>Sphaerophoria scripta</i> (Linnaeus, 1758)	9	Mainly ground layer aphids (90) Found on a wide range of plant species	Typically multiple generations (>2/year in Europe) (23)	5–7 months in Europe	Apparently puparium in grass roots	Wide range of habitats, including agro-ecosystems	Average of 365 eggs/female in the lab (20–22°C) (76)	Highly
<i>Calliphora vicina</i> (Robineau-Desvoidy, 1830)	8	Feed on various carrion and processed meats (22)	Egg to adult in 15–22 days at 10°C (18) Multiple generations/year, but dependent on weather (1) Adult longevity is 93–128 days (79)	Most abundant in the spring and summer months, but also occurs in winter (49)	Overwinters as larvae, pupae, or adults (1)	Occurs in common urban habitats (52) and woodland (42) and across all continents but Antarctica (19)	Approximately 2,000 eggs/female (88)	Not available

many differences in their biology, and it is important to recognize that there are therefore different challenges and opportunities in how they can be managed for crop pollination outcomes. There is also a great diversity within each of these clades; thus, while we can make generalizations, each crop and potential pollinator will have its own particularities.

Bees depend entirely on floral resources for both larval and adult stages, while most flies and many other non-bee flower visitors do not. Of the non-bees identified as commonly visiting crops in this review, adults primarily use floral resources (and then mostly nectar), whereas non-bee larval lifestyles are widely variable, even within families. Syrphidae are dominated, but not exclusively so, by species with either predacious larvae (consuming aphids and other small soft-bodied insects) or larvae living in aquatic or semiaquatic environments feeding on a broad range of organic material (**Table 2**). The larvae of many Calliphoridae feed on a range of carrion but also include species parasitic on earthworms (*Pollenia* spp.) (101) and those that feed on the larvae and eggs of other insects (*Stomorrhina* spp.) (112). The larger flies that typically visit the flowers of many crops have larvae that feed on resources that are common in agricultural landscapes—aphids, dung, and decaying organic matter (**Table 2**). Lepidopterans often have specific host–plant relationships for larval feeding, and as a consequence, agricultural intensification at landscape scales often favors species with more general host–plant larval feeding requirements (70). Flower-visiting beetles also have a wide variety of larval food preferences, reflecting the high taxonomic diversity of flower-visiting taxa (58, 67), and several taxa are known to use fruit as food and/or brood sites (57, 103).

Many non-bee insect pollinators, such as flies, butterflies, and beetles, are capable of large flight distances relative to most bees. This is in part because non-bee insects do not provision young and maintain nests, as do bees (the exception being some non-bee hymenopterans). Because bees provision their young with pollen, the larvae remain in the nest or colony, and the adult bees have an attachment that limits their freedom to move. This attachment is strongest for the social bees with perennial colonies, but even for solitary bees, adult females return to the nest repeatedly during establishment (74). Among the bees, only some larger-bodied species are known to travel great distances (e.g., 50 km for Euglossine bees) (82), while most have a foraging range under 1 km (33, 38, 95, 113). In contrast, long-distance flights of between 50 and 110 km have been recorded for Syrphid flies, including *Episyrphus balteatus* and *Sphaerophoria scripta* (2, 34). Such travel distances may be important with regard to the role of non-bee insects in transgene flow (83).

Dipteran pollinators, in particular Calliphoridae and Syrphidae, are also capable of quickly responding to changes in resource availability through high mobility, high fecundity, and short life cycles. Hoverflies, for example, are capable of producing several generations per year (**Table 2**). As a consequence, some flies that visit many crops produce multiple generations per year and produce eggs numbering in the thousands per female (**Table 2**); bees will typically have fewer generations per year and fewer offspring produced (26, 74). As bees need to provision their larvae with pollen, they generally invest in fewer but relatively larger eggs compared with flies. At the extreme, some *Xyllocopa* produce eggs that are approximately half the length of the adult female's body (74). These different tendencies in reproductive strategy would generally be expected to permit the flies to have faster population growth rates than bees when conditions are suitable.

While there are indeed species of social bees (particularly *Apis* spp.) that share information on foraging opportunities in the colony and then recruit to those resources in large numbers, this level of social behavior is not common among bees, and in fact, the majority of bee species are solitary (i.e., no social structure) and therefore do not share information (21, 74, 87). Thus, while the social bees have higher reproductive potential than other bees and many flies if compared on an individual-to-individual basis, the performance of the queen needs to be judged relative to the whole colony's labors that support her.

Bees and flies also exhibit marked differences in flower foraging behavior (54). These differences are particularly important in hybrid seed crops, which require movement between rows of intermixed plantings of outcrossing crops. For example, in vegetable hybrid seed crops, both honey bees and wild bees often show preferences for one line over the other (50) and do not frequently move between the lines (66). Non-bees have been found to move between male and female lines in vegetable hybrid seed production systems more regularly than do bees (29).

Many flies have worldwide distributions, including the major crop-visiting fly families and the five fly species that most commonly visit crops (**Table 2**), suggesting that they are well adapted to transformed landscapes. *Eristalis tenax* originated in Europe or the Middle East and now has a global distribution, a history shared with many other imported crop pollinator taxa, such as houseflies (*Musca domestica*) and some blowflies (*Calliphora* spp.). This has facilitated their expansion into many agricultural and urban environments, where they have been recorded as flower visitors and pollinators (41, 98).

Responses of bee and non-bee pollinators to agricultural environments are complex and understudied (92), and generalized patterns that hold across multiple pollinator taxa and crop types are difficult to identify (4, 45). Nonetheless, different insect taxa respond differently to changes in surrounding land use (62, 97). Landscape intensification filters pollinators by functional traits favoring generalist feeding habits and larger body sizes (30). For bees, there is strong evidence to suggest that richness and visitation rates to crops decline with increasing distance from natural habitats (86), likely due to nesting and nutritional habitat needs that are limited in agricultural fields (110). For example, many solitary bee species are ground nesters (74) and can thus be impacted by farm management methods such as frequent tillage (104), while a few more tolerant bee species, such as some *Lasioglossum* spp. (27, 40), remain in highly transformed landscapes. In contrast, hoverfly diversity appears to be less negatively impacted by changes in land use than diversity of other taxa, and many hoverfly species have been found to benefit from crop production (55). Some studies find that hoverfly assemblages maintain species richness and abundance away from remnant and seminatural vegetation, in contrast to wild bees.

In some agricultural systems, specific habitat patches have been shown to be important for non-bee pollinators. Proximity to rainforest influenced the assemblages of pollinating beetles that visited atemoya flowers in tropical Australia (5). Increased hedgerow cover at the landscape level was associated with greater hoverfly visitation rates and enhanced pollination of plants, as well as reduced abundance of crop pests (17). However, hedgerows can also act as a barrier to pollinator movement (60).

KNOWLEDGE GAPS AND RESEARCH PRIORITIES

Nocturnal Pollination

Despite nocturnal pollination occurring in plant species from 30% of angiosperm families (7), the contribution of nocturnal animals to crop pollination has rarely been examined (16, 64). The role of moths may be particularly underestimated. They are considered the most speciose order of flower-visiting insects (approximately 123,000 species versus 17,000 bee species; 108) and are considered pollinators of at least 289 plant species (69). Crops for which nocturnal pollinators have been recognized as important include papaya (moths) (72), *Agave tequilana* (bats) (102) and durian (bats and nocturnal bees) (109). Each of these crops has floral characteristics expressed more strongly at night, e.g., papaya flowers open at dusk and are typically more strongly scented at night than during the day (20); paniculate agaves (e.g., *A. tequilana*) typically produce large quantities of nectar and pollen at night (94); durian anthers dehisce in the evening, pollen viability then steadily decreases (109), and floral structures abscise by morning (47).

Nocturnal pollination may also be more important than is generally recognized even for crops with prominent daytime pollinators. Moths carry pollen that suggests that they are at least flower visitors of various crops, including *Brassica/Raphanus* (69), and nocturnal pollination may support fruit set in lowbush blueberry *Vaccinium angustifolium* (16). Moths are also suggested to be pollinators of *Lonicera caerulea* (Haskap) cultivars, as some flowers open in the evening and are still open in the morning (28).

For those crops that allow pollination across the day–night cycle, nocturnal pollinators might prove to be significant as a compliment to the services of diurnal pollinators. However, the paucity of studies assessing nocturnal pollination in crops limits our ability to develop strategies for growers to manage pollinators to optimize nocturnal pollination.

The Importance of Small Non-Bee Insects as Crop Pollinators

Small non-bee insects (defined as having body length smaller than 3 mm) are also key pollinators of a small number of crops, including cocoa (*T. cacao* L.), pollinated by *Forcipomyia* midges (37); fig (*Ficus* spp.), of which some varieties still depend on pollination by fig wasps (subfamily: Agaonidae) (14); and jackfruit (*A. heterophyllum* Lam.), pollinated by the gall midge *Clinodiplosis ultracrepidata* Gagné (Cecidomyiidae) (31). The flowers and inflorescences of both figs and jackfruit also support the larval stages of their pollinators. Fig wasps oviposit within the inflorescence (syconium), where the larvae then develop (89), while fungus-infected male flowers of jackfruit provide a brood site and larval nutrition for their midge pollinators (31). Thrips (Thysanoptera) have been implicated in the pollination of 24 crop species (59). In other crops, small non-bee insects were not found to contribute significantly to pollination. For example, while studies have reported high abundances and diversity of small insects (particularly Diptera and Thysanoptera) within flowering crops, including pak choi (*Brassica rapa* ssp. *chinensis*) (51), onion (*Allium cepa*) (106), and carrot (*Daucus carota*) (6), they were not found to contribute significantly to pollination. For some of the crops where small non-bee pollinators are thought to play an important role, habitat management is important to support high abundances in orchards. For example, management strategies to support *Forcipomyia* midges in cocoa include the provision of moist, decaying organic material within orchards, the provision of shade, and the minimal use of insecticides (narrow rather than broad spectrum) (12). Overall, we have little knowledge of the species of small non-bee insects that visit crops, apart from a few key crop species, and little to no knowledge of their effectiveness in most crops.

Limited Data Availability: Pollinator Efficiency and Gene Flow

Most records of non-bees visiting crops are no more than observations of insects visiting flowers. Few studies provide data relating to pollen deposition or fruit or seed set estimates after visitation by specific taxa (Table 1). While we know that quantity and quality of pollen transferred varies among taxa (34, 84), this area deserves significant further research. Both bee and non-bee insects also act as significant flower visitors and pollinators of many wild and cultivated plants that are generally assumed to be wind pollinated or autogamous, such as rice (83). Insect visitors to these crops can affect gene flow even if they have little influence on overall fruit set, so they deserve more attention, particularly in this context.

MANAGEMENT OF NON-BEE POLLINATORS FOR AGRICULTURE

Given the roles for non-bee pollinators indicated by our review, we advocate the use of two broad categories for non-bee pollinator management: (a) management for generalist-pollinated crops,

dominated by medium to large flies, and (b) management for crops with more specialized associations with flower visitors, such as beetles, thrips, and small fly pollinators.

As floral resources are limited in many agricultural habitats, schemes encouraging wildflower strips have been implemented in many parts of the world to enhance biodiversity and connectivity in agricultural landscapes (3, 24, 48, 65). Identifying suitability of different flower resources is important because studies have shown that, even when many different flowers are available, bees do not collect pollen from all species (39). While it is well established that bee colonies require a balanced diet with high protein for optimal larval development (9, 43), and floral richness has been found to be important to hoverflies at the local scale (93), we know relatively little about floral resource needs of non-bee flower visitors.

Many non-bee flower visitors across several different orders have partially aquatic life histories (85) including *E. tenax*—one of the most important non-bee pollinators globally. *E. tenax* and other members of the Eristalinae subfamily of the Syrphidae begin their life feeding on decaying organic matter in semiaquatic habitats (96). Agricultural habitats that provide resources for the development of these larvae include ponds (99); effluent holding pits; and streams containing sewage, decaying vegetation, or carcasses (80). Syrphid abundance has been specifically associated with pond habitats in agricultural landscapes and strawberry pollination (99). The effective management and conservation of these habitats is thus an important component of management strategies to cater to non-bee pollination services.

Management practices in the broader landscape can also be modified to promote particular taxa. Less intensively managed grasslands are associated with higher syrphid abundance and beetle species richness, perhaps in response to increases in plant height and leaf litter (93). Similarly, management practices such as delayed summer cutting of grasslands to provide extended flower forage resources and providing uncut refuge areas have been found to promote hoverfly abundance and diversity (73), and staggered mowing of road verges to leave these habitats undisturbed when they are especially productive promotes abundance of bee and non-bee pollinators (13). The diversity and abundance of moths have been shown to decline with grassland intensification (increased fertilizer use, grazing, and mowing), although host generalists were less affected; the structure of the moth community depended in part on whether the grassland was animal grazed or mowed (70). As well as diversification of pastures and grasslands, non-bee pollinators are likely to respond positively to mixing and alternating grazing intensity practices at the local and landscape levels. At the landscape scale, however, floral resource effects on syrphids might be moderated by availability of prey for the predaceous species (91). It may also be that the high mobility of some syrphids, such as *E. tenax*, allows populations to exploit widely scattered resources (71).

One of the most promising benefits of integrating non-bees into agricultural cropping systems is the potential for synergies across other ecosystem services, such as biological control of pests (10, 56). One of the major groups of the Syrphidae—the Syrphinae—are both pollinators as adults and predatory at larval stages on aphids and other major crop insect pests (44). In times of few floral resources, honeydew from aphids may also be utilized as an additional energy source by adult hoverflies (105). Other insect taxa that have dual roles as crop pollinators and pest predators include wasps and tachinid flies (53, 62) if flowering phenology is matched with pest egg availability (75). These complementary services have the potential to reduce the cost to growers by reducing the need for pesticide applications, which may reduce pressure on other beneficial insect pollinator groups utilizing the local environment (81).

Finally, careful planning needs to accompany decisions and strategies to promote non-bee pollinators, as some pollinating fly species can spread disease and negatively impact other types of agricultural production, such as livestock (e.g., blowfly strike; 100). Nitidulidae beetles may be

fruit pests, and many flower-visiting butterflies, e.g., cabbage whites, are also crop pests. Identifying the trade-offs and identifying effective management options under these conditions are important first steps.

In conclusion, non-bee taxa are important contributors to many global food crops, yet significant knowledge gaps exist with respect to the identity and efficiency of these taxa. These limitations make it difficult to target the management of specific pollinators for specific crops or to estimate the economic value of these taxa to crop production worldwide.

The diversity of resources required for non-bee as well as bee reproduction means that farm-level as well as landscape management is critical to close the yield gap and create sustainable agricultural systems and communities. Targeting specific pollinators for specific crops will require further research evaluating the efficiency and life history needs of diverse taxa and how these vary in time and with the scale of habitat quality.

DISCLOSURE STATEMENT

The authors are not aware of any affiliations, memberships, funding, or financial holdings that might be perceived as affecting the objectivity of this review.

ACKNOWLEDGMENTS

The authors thank Jeremy Jones for conducting literature searches and assisting with data manipulation, Emma Goodwin for generating figures, and Warrick Nelson and Manu Saunders for commenting on an earlier version of the manuscript. R.R. was funded by Australian Research Council Discovery Early Career Researcher Award DE170101349.

LITERATURE CITED

1. Aak A, Birkemoe T, Leinaas HP. 2011. Phenology and life history of the blowfly *Calliphora vicina* in stockfish production areas. *Entomol. Exp. Appl.* 139:35–46
2. Aubert J, Goeldlin de Tiefenau P. 1981. Observations sur les migrations de Syrphides (Dipt.) dans les Alpes de Suisse occidentale. *Mitt. Schweiz. Entomol. Ges. Bull. Soc. Entomol. Suisse* 54:377–81
3. Balzan MV, Bocci G, Moonen A-C. 2014. Augmenting flower trait diversity in wildflower strips to optimise the conservation of arthropod functional groups for multiple agroecosystem services. *J. Insect Conserv.* 18:713–28
4. Bartomeus I, Cariveau DP, Harrison T, Winfree R. 2018. On the inconsistency of pollinator species traits for predicting either response to land-use change or functional contribution. *Oikos* 127:306–15
5. Blanche R, Cunningham SA. 2005. Rain forest provides pollinating beetles for atemoya crops. *J. Econ. Entomol.* 98:1193–201
6. Bohart GE, Nye WP. 1960. *Insect pollinators of carrots in Utah*. Bull. 419, Agric. Exp. Stn., Utah State Univ., Logan
7. Borges RM, Somanathan H, Kelber A. 2016. Patterns and processes in nocturnal and crepuscular pollination services. *Q. Rev. Biol.* 91:389–418
8. Brittain C, Williams N, Kremen C, Klein AM. 2013. Synergistic effects of non-*Apis* bees and honey bees for pollination services. *Proc. R. Soc. B* 280:20122767
9. Bukovinszky T, Rikken I, Evers S, Wäckers F, Biesmeijer J, et al. 2016. Effects of pollen species composition on the foraging behaviour and offspring performance of the mason bee *Osmia bicornis* (L.). *Basic Appl. Ecol.* 18:21–30
10. Campbell AJ, Wilby A, Sutton P, Wäckers F. 2017. Getting more power from your flowers: multi-functional flower strips enhance pollinators and pest control agents in apple orchards. *Insects* 8:E101

11. Cik Mohd Rizuan ZA, Noor Hisham H, Samsudin A. 2013. *Role of pollinating weevil (Elaeidobius kamerunicus), seasonal effect and its relation to fruit set in oil palm area of FELDA Sababat*. Paper presented at PIPOC 2013 Conf., Kuala Lumpur, Malaysia, Nov. 19–21
12. Claus G, Vanhove W, Van Damme P, Smagghe G. 2018. Challenges in cocoa pollination: the case of Côte d'Ivoire. In *Pollination in Plants*, ed. PW Mokwala, art. 4. London: IntechOpen
13. Cole LJ, Brocklehurst S, Robertson D, Harrison W, McCracken DI. 2017. Exploring the interactions between resource availability and the utilisation of semi-natural habitats by insect pollinators in an intensive agricultural landscape. *Agric. Ecosyst. Environ.* 246:157–67
14. Cook J, Rasplus J-Y. 2003. Mutualists with attitude: coevolving fig wasps and figs. *Trends Ecol. Evol.* 18:241–48
15. Córdoba SA, Cocucci AA. 2011. Flower power: its association with bee power and floral functional morphology in papilionate legumes. *Ann. Bot.* 108:919–31
16. Cutler G, Reeh KW, Sproule JM, Ramanaidu K. 2012. Berry unexpected: nocturnal pollination of low-bush blueberry. *Can. J. Plant. Sci.* 92:707–11
17. Dainese M, Montecchiari S, Sitzia T, Sigura M, Marini L. 2017. High cover of hedgerows in the landscape supports multiple ecosystem services in Mediterranean cereal fields. *J. Appl. Ecol.* 54:380–88
18. Davies L, Ratcliffe GG. 1994. Development rates of some pre-adult stages in blowflies with reference to low temperatures. *Med. Vet. Entomol.* 8:245–54
19. Dear JP. 1985. *Calliphoridae (Insecta: Diptera)*. Fauna N. Z. 8. Wellington, N. Z.: Sci. Inform. Cent.
20. Dey K, Mondal S, Mandal S. 2016. Flower visitor diversity with reference to pollen dispersal and pollination of *Carica papaya* L. *Int. J. Adv. Res. Biol. Sci.* 3:65
21. Donovan BJ. 2007. *Apoidea (Insecta: Hymenoptera)*. Fauna N. Z. 57. Christchurch, N. Z.: Landcare Res.
22. Donovan SE, Hall MJ, Turner BD, Moncrieff CB. 2006. Larval growth rates of the blowfly, *Calliphora vicina*, over a range of temperatures. *Med. Vet. Entomol.* 20:106–14
23. Dziock F. 2006. Life-history data in bioindication procedures, using the example of hoverflies (Diptera, syrphidae) in the Elbe floodplain. *Int. Rev. Hydrobiol.* 91:341–63
24. Ekroos J, Olsson O, Rundlöf M, Wätzold F, Smith HG. 2014. Optimizing agri-environment schemes for biodiversity, ecosystem services or both? *Biol. Conserv.* 172:65–71
25. Forbes S, Northfield T. 2016. Increased pollinator habitat enhances cacao fruit set and predator conservation. *Ecol. Appl.* 27:887–99, R, PJ.
26. Forrest JRK, Cross R, CaraDonna PJ. 2019. Two-year bee, or not two-year bee? How voltinism is affected by temperature and season length in a high-elevation solitary bee. *Am. Nat.* 193:560–74
27. Forrest JRK, Thorp RW, Kremen C, Williams NM. 2015. Contrasting patterns in species and functional-trait diversity of bees in an agricultural landscape. *J. Appl. Ecol.* 52:706–15
28. Frier SD, Somers CM, Sheffield CS. 2016. Floral longevity, nectar production, pollen release, and stigma receptivity in Haskap (*Lonicera caerulea*). *J. Pollinat. Ecol.* 19:81–87
29. Gaffney A, Bohman B, Quarrell SR, Brown PH, Allen GR. 2018. Frequent insect visitors are not always pollen carriers in hybrid carrot pollination. *Insects* 9:61
30. Gámez-Virués S, Perović DJ, Gossner MM, Börschig C, Blüthgen N, et al. 2015. Landscape simplification filters species traits and drives biotic homogenization. *Nat. Commun.* 6:8568
31. Gardner EM, Gagné RJ, Kendra PE, Montgomery WS, Raguso RA, et al. 2018. A flower in fruit's clothing: pollination of jackfruit (*Artocarpus heterophyllus*, Moraceae) by a new species of gall midge, *Clinodiplosis ultracrepidata* sp. nov. (Diptera: Cecidomyiidae). *Int. J. Plant Sci.* 179:350–67
32. Garibaldi LA, Steffan-Dewenter I, Winfree R, Aizen MA, Bommarco R, et al. 2013. Wild pollinators enhance fruit set of crops regardless of honey bee abundance. *Science* 339:1608–11
33. Gathmann A, Tscharrntke T. 2002. Foraging ranges of solitary bees. *J. Anim. Ecol.* 71:757–64
34. Gatter W, Schmid U. 1990. Wanderungen der Schwebfliegen (Dipter, Syrphidae) am Randecker Maar. *Spixiana* 15:1–100
35. Geusenpfister H. 1987. Studies on the biology and reproductive capacity of *Episyrphus balteatus* Deg. (Dipt., Syrphidae) under greenhouse conditions. *J. Appl. Entomol.* 104:261–70
36. Ghorpade KD. 1981. Insect prey of syrphids (Diptera) from India and neighbouring countries: a review and bibliography. *Trop. Pest Manag.* 27:62–82

37. Glendinning DR. 1972. Natural pollination of cocoa. *New Phytol.* 71:719–29
38. Greenleaf SS, Williams NM, Winfree R, Kremen C. 2007. Bee foraging ranges and their relationship to body size. *Oecologia* 153:589–96
39. Gresty CEA, Clare E, Devey DS, Cowan RS, Csiba L, et al. 2018. Flower preferences and pollen transport networks for cavity-nesting solitary bees: implications for the design of agri-environment schemes. *Ecol. Evol.* 8:7574–87
40. Hall MA, Nimmo DG, Cunningham SA, Walker K, Bennett AF. 2019. The response of wild bees to tree cover and rural land use is mediated by species' traits. *Biol. Conserv.* 231:1–12
41. Hennig EI, Ghazoul J. 2012. Pollinating animals in the urban environment. *Urban Ecosyst.* 15:149–66
42. Henning J, Schnitzler FR, Pfeiffer DU, Davies P. 2005. Influence of weather conditions on fly abundance and its implications for transmission of rabbit haemorrhagic disease virus in the North Island of NZ. *Med. Vet. Entomol.* 19:251–62
43. Höcherl N, Siede R, Illies I, Gätschenberger H, Tautz J. 2011. Evaluation of the nutritive value of maize for honey bees. *J. Insect Physiol.* 58:278–85
44. Hodgkiss D, Brown MJF, Fountain MT. 2018. Syrphine hoverflies are effective pollinators of commercial strawberry. *J. Pollinat. Ecol.* 22:55–66
45. Holzschuh A, Dainese M, González-Varo JP, Mudri-Stojnić S, Riedinger V, et al. 2016. Mass-flowering crops dilute pollinator abundance in agricultural landscapes across Europe. *Ecol. Lett.* 19:1228–36
46. Hong BM, Hung HQ. 2010. Effect of temperature and diet on the life cycle and predatory capacity of *Episyrphus balteatus* (De Geer) (Syrphidae: Diptera) cultured on *Aphis gossypii* (Glover). *J. Int. Soc. Southeast Asian Agric. Sci.* 16:98–103
47. Honsho C, Somsri T, Tetsumura T, Yamashita K, Yonemori K. 2007. Effective pollination period in durian (*Durio zibethinus* Murr.) and the factors regulating it. *Sci. Horticul.* 111:193–96
48. Howlett BG, Davidson MM, Mathers D, Pyke NB. 2013. Hedgerow plants to support crop pollination and pest management. *Weta* 46:3–12
49. Howlett BG, Davidson MM, Pattermore DE, Walker MK, Nelson WR. 2016. Seasonality of calliphorid and sarcophagid flies across Canterbury arable farms requiring pollinators. *N.Z. Plant Protect.* 69:290–95
50. Howlett BG, Lankin-Vega GO, Pattermore DE. 2015. Native and introduced bee abundances on carrot seed crops in New Zealand. *N. Z. Plant Prot.* 68:373–79
51. Howlett BG, Walker MK, McCallum JA, Teulon DAJ. 2009. Small flower-visiting arthropods in New Zealand pak choi fields. *N. Z. Plant Prot.* 62:86–91
52. Hwang C, Turner BD. 2005. Spatial and temporal variability of necrophagous Diptera from urban to rural areas. *Med. Vet. Entomol.* 19:379–91
53. Inclán DJ, Dainese M, Cerretti P, Paniccía D, Marini L. 2016. Spillover of tachinids and hoverflies from different field margins. *Basic Appl. Ecol.* 17:33–42
54. Inouye DW, Larson BMH, Ssymank A, Kevan PG. 2015. Flies and flowers III: ecology of foraging and pollination. *J. Pollinat. Ecol.* 16:115–33
55. Jauker F, Wolters V. 2008. Hover flies are efficient pollinators of oilseed rape. *Oecologia* 156:819–23
56. Joshi S, Ballal CR. 2013. Syrphid predators for biological control of aphids. *J. Biol. Control* 27:151–70
57. Jürgens A, Webber AC, Gottsberger G. 2000. Floral scent compounds of Amazonian Annonaceae species pollinated by small beetles and thrips. *Phytochemistry* 55:551–58
58. Kevan PG. 1983. Insects as flower visitors and pollinators. *Annu. Rev. Entomol.* 28:407–53
59. Kirk W. 1997. Distribution, abundance, and population dynamics. In *Thrips as Crop Pests*, ed. T Lewis, pp. 217–58. Wallingford, UK: CAB Int.
60. Klaus F, Bass J, Marholt L, Müller B, Klatt B, Kormann U. 2015. Hedgerows have a barrier effect and channel pollinator movement in the agricultural landscape. *J. Landscape Ecol.* 8:22–31
61. Kleijn D, Winfree R, Bartomeus I, Carvalheiro LG, Henry M, et al. 2015. Delivery of crop pollination services is an insufficient argument for wild pollinator conservation. *Nat. Commun.* 6:7414
62. Klein A-M, Steffan-Dewenter I, Buchori D, Tschardt T. 2002. Effects of land-use intensity in tropical agroforestry systems on coffee flower-visiting and trap-nesting bees and wasps. *Conserv. Biol.* 16:1003–14
63. Klein A-M, Vaissiere BE, Cane JH, Steffan-Dewenter I, Cunningham SA, et al. 2007. Importance of pollinators in changing landscapes for world crops. *Proc. R. Soc. B* 274:303–13

64. Knop E, Gerpe C, Ryser R, Hofmann F, Menz MHM, et al. 2018. Rush hours in flower visitors over a day–night cycle. *Insect Conserv. Divers.* 11:267–75
65. Knop EVA, Kleijn D, Herzog F, Schmid B. 2006. Effectiveness of the Swiss agri-environment scheme in promoting biodiversity. *J. Appl. Ecol.* 43:120–27
66. Kobayashi K, Tsukamoto S, Tanaka A, Niikura S, Ohsawa R. 2010. Selective flower visitation behavior by pollinators in a radish F1 seed production field. *Breeding Sci.* 60:203–11
67. Krenn HW, Plant JD, Szucsich NU. 2005. Mouthparts of flower-visiting insects. *Arthropod Struct. Dev.* 34:1–40
68. Lim T, Luders L. 2008. Durian flowering, pollination and incompatibility studies. *Ann. Appl. Biol.* 132:151–65
69. Macgregor CJ, Pocock MJO, Fox R, Evans DM. 2015. Pollination by nocturnal Lepidoptera, and the effects of light pollution: a review. *Ecol. Entomol.* 40:187–98
70. Mangels J, Fiedler K, Schneider FD, Blüthgen N. 2017. Diversity and trait composition of moths respond to land-use intensification in grasslands: Generalists replace specialists. *Biodivers. Conserv.* 26:3385–405
71. Marcetic LF, Ludoški J, Milankov V. 2013. Phenotypic diversity and landscape genetics of *Eristalis tenax* in a spatially heterogeneous environment, Durmitor Mountain (Montenegro). *Ann. Zool. Fenn.* 50:262–78
72. Martins DJ, Johnson SD. 2009. Distance and quality of natural habitat influence hawkmoth pollination of cultivated papaya. *Int. J. Trop. Insect Sci.* 29:114–23
73. Meyer S, Unternährer D, Arlettaz R, Humbert J-Y, Menz MHM. 2017. Promoting diverse communities of wild bees and hoverflies requires a landscape approach to managing meadows. *Agric. Ecosyst. Environ.* 239:376–84
74. Michener CD. 2007. *The Bees of the World*. Baltimore, MD: Johns Hopkins Univ. Press
75. Morris MC, Li FY. 2000. Coriander (*Coriandrum sativum*) “companion plants” can attract hoverflies, and may reduce pest infestation in cabbages. *N. Z. J. Crop Hortic. Sci.* 28:213–17
76. Nengel S, Drescher W. 1991. Studies on the biology of *Sphaerophoria scripta* L. (Diptera, Syrphidae). *Acta Hortic. Sci.* 288:98–102
77. Nicholas S, Thyselius M, Holden M, Nordström K. 2018. Rearing and long-term maintenance of *Eristalis tenax* hoverflies for research studies. *J. Vis. Exp.* 135:e57711
78. Ollerton J, Winfree R, Tarrant S. 2011. How many flowering plants are pollinated by animals? *Oikos* 120:321–26
79. Perez C, Segura NA, Patarroyo MA, Bello FJ. 2016. Evaluating the biological cycle and reproductive and population parameters of *Calliphora vicina* (Diptera: Calliphoridae) reared on three different diets. *J. Med. Entomol.* 53:1268–75
80. Perez-Bañón C, Hurtado P, García-Gras E, Rojo S. 2013. SEM studies on immature stages of the drone flies (Diptera, Syrphidae): *Eristalis similis* (Fallen, 1817) and *Eristalis tenax* (Linnaeus, 1758). *Microsc. Res. Technol.* 76:853–61
81. Pineda A, Marcos-García MÁ. 2008. Use of selected flowering plants in greenhouses to enhance aphidophagous hoverfly populations (Diptera: Syrphidae). *Ann. Soc. Entomol.* 44:487–92
82. Pokorný T, Loose D, Dyker G, Quezada-Euán JJG, Eltz T. 2015. Dispersal ability of male orchid bees and direct evidence for long-range flights. *Apidologie* 46:224–37
83. Pu D, Shi M, Wu Q, Gao M, Liu J, et al. 2014. Flower-visiting insects and their potential impact on transgene flow in rice. *J. Appl. Ecol.* 51:1357–65
84. Rader R, Bartomeus I, Garibaldi LA, Garratt MPD, Howlett BG, et al. 2016. Non-bee insects are important contributors to global crop pollination. *PNAS* 113:146–51
85. Raitif J, Plantegenest M, Roussel J-M. 2019. From stream to land: ecosystem services provided by stream insects to agriculture. *Agric. Ecosyst. Environ.* 270–71:32–40
86. Ricketts TH, Regetz J, Steffan-Dewenter I, Cunningham SA, Kremen C, et al. 2008. Landscape effects on crop pollination services: Are there general patterns? *Ecol. Lett.* 11:1121
87. Robinson GE. 1992. Regulation of division of labor in insect societies. *Annu. Rev. Entomol.* 37:637–65

88. Rognes K. 1991. *Blowflies (Diptera, Calliphoridae) of Fennoscandia and Denmark*. Fauna Entomol. Scand. 24. Leiden, Neth.: E. Brill
89. Rønsted N, Weiblen GD, Cook JM, Salamin N, Machado CA, Savolainen V. 2006. 60 million years of co-divergence in the fig–wasp symbiosis. *Proc. R. Soc. B* 272:2593–99
90. Rotheray GE. 1993. Colour guide to hoverfly larvae (Diptera: Syrphidae) in Britain and Europe. *Dipterists Digest* 9:1345409
91. Schirmel J, Albrecht M, Bauer P-M, Sutter L, Pfister SC, Entling MH. 2018. Landscape complexity promotes hoverflies across different types of semi-natural habitats in farmland. *J. Appl. Ecol.* 55:1747–58
92. Senapathi D, Goddard MA, Kunin WE, Baldock KCR. 2017. Landscape impacts on pollinator communities in temperate systems: evidence and knowledge gaps. *Funct. Ecol.* 31:26–37
93. Sjödin NE, Bengtsson J, Eklom B. 2008. The influence of grazing intensity and landscape composition on the diversity and abundance of flower-visiting insects. *J. Appl. Ecol.* 45:763–72
94. Slauson L. 2000. Pollination biology of two chiropterophilous agaves in Arizona. *Am. J. Bot.* 87:825–36
95. Smith JP, Heard TA, Beekman M, Gloag R. 2017. Flight range of the Australian stingless bee *Tetragonula carbonaria* (Hymenoptera: Apidae). *Austral. Entomol.* 56:50–53
96. Speight MCD. 2011. *Species Accounts of European Syrphidae (Diptera)*, 2014. Dublin: Syrph Net Publ.
97. Stanley DA, Stout JC. 2013. Quantifying the impacts of bioenergy crops on pollinating insect abundance and diversity: a field-scale evaluation reveals taxon-specific responses. *J. Appl. Ecol.* 50:335–44
98. Stavert JR, Pattemore DE, Bartomeus I, Gaskett AC, Beggs JR, Diekötter T. 2018. Exotic flies maintain pollination services as native pollinators decline with agricultural expansion. *J. Appl. Ecol.* 55:1737–46
99. Stewart RIA, Andersson GKS, Brönmark C, Klatt BK, Hansson L-A, et al. 2017. Ecosystem services across the aquatic–terrestrial boundary: linking ponds to pollination. *Basic Appl. Ecol.* 18:13–20
100. Tellam RL, Bowles VM. 1997. Control of blowfly strike in sheep: current strategies and future prospects. *Int. J. Parasitol.* 27:261–73
101. Thomson AJ, Davies DM. 1973. The biology of *Pollenia rudis*, the cluster fly (Diptera: Calliphoridae): I. Host location by first-instar larvae. *Can. Entomol.* 105:335–41
102. Trejo-Salazar R-E, Eguiarte LE, Suro-Piñera D, Medellín RA. 2016. Save our bats, save our tequila: industry and science join forces to help bats and agaves. *Nat. Areas J.* 36:523–30
103. Tuo Y, Koua HK, Hala N. 2011. Biology of *Elaeidobius kamerunicus* and *Elaeidobius plagiatus* (Coleoptera: Curculionidae) main pollinators of oil palm in West Africa. *Eur. J. Sci. Res.* 49:426–32
104. Ullmann KS, Meisner MH, Williams NM. 2016. Impact of tillage on the crop pollinating, ground-nesting bee, *Peponapis pruinosa* in California. *Agric. Ecosyst. Environ.* 232:240–46
105. van Rijn PCJ, Kooijman J, Wäckers FL. 2013. The contribution of floral resources and honeydew to the performance of predatory hoverflies (Diptera: Syrphidae). *Biol. Control* 67:32–38
106. Walker MK, Howlett BG, Wallace AR, McCallum JA, Teulon DAJ. 2011. The diversity and abundance of small arthropods in onion, *Allium cepa*, seed crops, and their potential role in pollination. *J. Insect Sci.* 11:98
107. Wang R, Aylwin R, Barwell L, Chen X-Y, Chen Y, et al. 2015. The fig wasp followers and colonists of a widely introduced fig tree, *Ficus microcarpa*. *Insect Conserv. Divers.* 8:322–36
108. Wardhaugh CW. 2015. How many species of arthropods visit flowers? *Arthropod-Plant Interact.* 9:547–65
109. Wayo K, Phankaew C, Stewart AB, Bumrungsri S. 2018. Bees are supplementary pollinators of self-compatible chiropterophilous durian. *J. Trop. Ecol.* 34:41–52
110. Williams NM, Crone EE, Roulston TH, Minckley RL, Packer L, Potts SG. 2010. Ecological and life history traits predict bee species responses to environmental disturbances. *Biol. Conserv.* 143:2280–91
111. Winfree R, Reilly JR, Bartomeus I, Cariveau DP, Williams NM, Gibbs J. 2018. Species turnover promotes the importance of bee diversity for crop pollination at regional scales. *Science* 359:791–93
112. Yeates D, Wiegmann B, eds. 2005. *The Evolutionary Biology of Flies*. New York: Columbia Univ. Press
113. Zurbuchen A, Landert L, Klaiber J, Müller A, Hein S, Dorn S. 2010. Maximum foraging ranges in solitary bees: Only few individuals have the capability to cover long foraging distances. *Biol. Conserv.* 143:669–76