

Annual Review of Entomology

Phoresy and Mites: More Than Just a Free Ride

Owen D. Seeman^{1,*} and David Evans Walter²

¹ Queensland Museum, South Brisbane, Queensland, Australia;
email: owen.seeman@qm.qld.gov.au

² University of the Sunshine Coast, School of Science, Technology and Engineering, Sippy Downs, Queensland, Australia

ANNUAL
REVIEWS **CONNECT**

www.annualreviews.org

- Download figures
- Navigate cited references
- Keyword search
- Explore related articles
- Share via email or social media

Annu. Rev. Entomol. 2023. 68:69–88

First published as a Review in Advance on
September 28, 2022

The *Annual Review of Entomology* is online at
ento.annualreviews.org

<https://doi.org/10.1146/annurev-ento-120220-013329>

Copyright © 2023 by the author(s). This work is licensed under a Creative Commons Attribution 4.0 International License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited. See credit lines of images or other third-party material in this article for license information.

*Corresponding author



Keywords

symbiosis, dispersal, parasitism, conditional relationships, host relationships, Acari

Abstract

Mites are masters at attaching to larger animals, often insects, in a temporary symbiosis called phoresy that allows these tiny animals to exploit patchy resources. In this article, we examine phoresy in the Acari, including those that feed on their carriers in transit, from a broad perspective. From a phylogenetic perspective, phoresy has evolved several times from free-living ancestors but also has been lost frequently. Rotting logs appear to be the first patchy resource exploited by phoretic mites, but the evolution of rapid life cycles later permitted exploitation of short-lived resources. As phoresy is a temporary symbiosis, most species have off-host interactions with their carrier. These relationships can be highly complex and context dependent but often are exploitative of the carrier's resources or progeny. Transitions from phoresy to parasitism seem widespread, but evidence for transitions from obligate phoretic parasitism to permanent parasitism is weak.

Deutonymph:

the third life stage of mites after the egg

Free living:

describes mites with no life stages that attach to carriers or hosts

Natal habitat:

the place where eggs are laid and most or all immature stages develop

1. INTRODUCTION

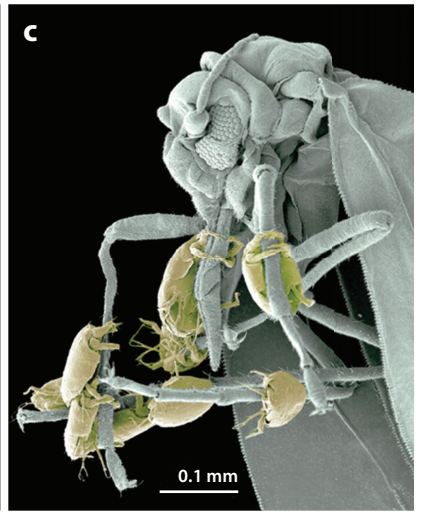
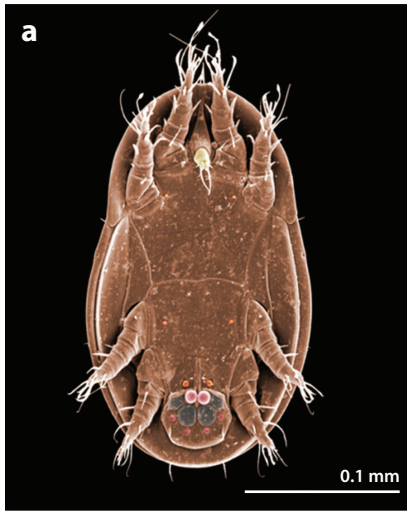
Mites include the smallest of all terrestrial arthropods, are the most diverse group of the Arachnida, and have infiltrated more habitats than the Insecta (142). Part and parcel of this acarine conquest and radiation is phoresy, their ability to hitch a ride on larger animals. Initially, phoresy was considered to include any relationship in which one animal transported another, even ants carrying their young; in which the relationship was temporary; and in which feeding on the carrier did not occur (82). However, Janet (55) struggled with the divide between phoresy and parasitism. *Antennophorus* species seem to be phoretic, but adult mites ride beneath the heads of ants, where they feed by deceiving their carrier into trophallaxis (40), while also being transported to new nests where their young develop as inquilines.

During the twentieth century, authors devised more restrictive definitions. Intraspecific transport was dropped, species that feed on the host continued to be excluded, and some definitions precluded development during transit—while also suggesting that movement should be from a less to a more suitable habitat (36). Since then, however, these definitions have increasingly broadened again. For the purposes of this review, we prefer the definitions of Camerik (21) and Walter & Proctor (142), with slight modification: Phoresy is a temporary symbiosis where one life stage of a smaller animal (the phoretic) attaches to another larger animal (the carrier) for dispersal. We emphasize that the symbiosis is temporary and usually interspecific. Thus, phoretics can have an additional interaction with a carrier; in this review, we discuss examples of mutualists, commensals, parasites, and parasitoids with a phoretic symbiosis within their life cycle. In addition, although the outcome of phoresy is dispersal, we do not exclude negative impacts on the carrier. Thus, impeding movement of the carrier or feeding from the host are still within the realm of a phoretic relationship.

Our preferred definition is, however, more restrictive than that of Bartlow & Agosta (7), which includes parasites such as ticks. Ticks may disperse to new habitats on hosts; however, this appears to be incidental, and we exclude these species. Our definition does include those parasites for which dispersal is integral to the relationship with the host, particularly those that are parasitic in only one life stage. This includes the highly diverse Parasitengona (larval parasites), several groups of Astigmata with parasitic deutonymphs, and groups such as the Heterozetionina and Paramesitridae that are parasitic as adults. We emphasize that phoresy is a symbiosis executed by one life stage. In mites, this is usually either a deutonymph or an adult (**Figure 1**). Exceptions are explicable. For instance, the flower-inhabiting mite *Hattena panopla* lives in flowers, and adults ride on visiting honeyeaters. As the duration of each flower is shorter than the life cycle of the mites, immature mites are also phoretic—but by riding on phoretic adult mites, a rare case of intraspecific phoresy (127).

In contrast, there is little question as to whether a mite is free living or phoretic. Free-living mites that occasionally appear on larger animals are termed accidental phoretics (1) or facultative phoretics (21). Accidental phoresy can be ecologically significant, as exemplified by the surprising number of ground-dwelling Oribatida found on birds (81) and beetles (100). Indeed, the earliest purported case of phoresy, from the Carboniferous Period (121), could be an accidental association between an oribatid and an insect.

Various authors have tried to classify phoretic relationships into subtypes (e.g., 132, 134, 144); the most enduring of these classifications is the division into facultative and obligate phoresy (1). The terms themselves are unfortunate, as phoretic species must, at some time, make an obligatory movement from the natal habitat. In this spirit, Camerik (21) redefined facultative phoresy as equivalent to accidental phoresy, while separating obligate phoretics into species that exhibit frequent obligate phoresy (i.e., in short-term habitats) and those exhibiting infrequent obligate phoresy (i.e., in long-term habitats).



(Caption appears on following page)

Figure 1 (Figure appears on preceding page)

Phoretic mites. (a,b) Astigmata: (a) phoretic deutonymph, ventral view; (b) on ologamasid mite venter. (c) Heterostigmata: *Polyphagotarsonemus latus* adults on *Bemisia tabaci*. (d) Parasitengona: *Arrenurus* water mite larvae on venter of *Tramea loewii*. (e) Parasitidae deutonymph (left) and Ereyetidae adults (right) on six-spined bark beetle, *Ips calligraphus*. (f) Mesostigmata, Trigynaspida: *Megisthanus thorelli* and Fedrizziidae adults on passalid beetle. (g) Mesostigmata, Uropodina: phoretic deutonymph, attached by anal pedicel. (h) Mesostigmata: *Poecilochirus* deutonymphs on *Nicrophorus* carrion beetle. Panels a–c copyright DE Walter. Panel d copyright Narelle Power and Damian White. Panel e copyright Clarence Holmes. Panel f copyright Anthony O'Toole. Panel g copyright Benjamin Fabian. Panel h copyright Jason Headley. All images used with permission.

It has been several decades since the last reviews of phoresy in various groups of Acari (3, 14, 31, 52, 53, 58, 103). The phylogeny of mites is now better resolved, allowing us to explore phoresy within a phylogenetic framework. We look for patterns within the major groups of phoretic mites, and across the Acari, before exploring three important aspects of phoresy: the impact on carriers, phoresy as part of a dynamic relationship between the mite and its carrier, and the evolution of other symbioses from phoresy.

2. THE ORIGINS OF PHORESY

Mites comprise two monophyletic superorders, Acariformes and Parasitiformes, which are of uncertain affinity to each other and other arachnid groups (66, 111). Associations with insects have formed multiple times in both groups, sometimes exhibiting highly specialized morphologies and phoretomorphic stages (142).

2.1. Acariformes

Acariform mites have a significant fossil record starting from the early Devonian Period (134). These early forms occurred mostly in soils, with the exception of Triassic records of plant-parasitic Eriophyoidea (125). None are obviously phoretic or otherwise symbiotic with insects until the Cretaceous Period. By this time, the major insect-associated lineages of acariform mites, Parasitengona (29, 73), Heterostigmata (61, 92, 93), and Astigmata (68) (**Figures 2** and **3**), are known from fossils.

These three major lineages have completely different associations with insects. Furthermore, several other groups have formed phoretic associations with insects, illustrating the propensity for mites to independently form phoretic relationships.

2.1.1. The Parasitengona. The Parasitengona comprise approximately 12,000 spp. that are almost all parasites as larvae and predators thereafter (38, 136). They include the chiggers (Trombiculoidea), which usually parasitize vertebrates; the water mites (Hydrachnida), which usually parasitize adult aquatic insects (**Figure 1d**); and several other families that parasitize terrestrial arthropods. The natal habitat is often not patchy or ephemeral, being a large body of water or a general habitat through which the mites search for prey. However, the free-living stages may occupy ephemeral habitats, such as temporary ponds (18) or nests (133). Thus, dispersal between these habitats as parasitic larvae is an essential part of their life cycle and possibly integral to the evolution of this unique life history (146).

2.1.2. The Heterostigmata. The Heterostigmata are a large, ecologically diverse group of mites with numerous associations with insects, including phoresy and permanent parasitism (58) (**Figure 2**). Early lineages associate closely with edaphic habitats and the nests of saproxylic social beetles, i.e., Passalidae and Scolytinae (Tarsocheylidae, Heterocheylidae). Beetles are common carriers, but the Pygmephoridae include numerous myrmecophiles (8). Fungivory predominates throughout the group, with adults traveling on insects between habitats rich with mycelia (48, 58).

Edaphic: associated with the soil

Saproxylic: feeding on dead or decaying wood

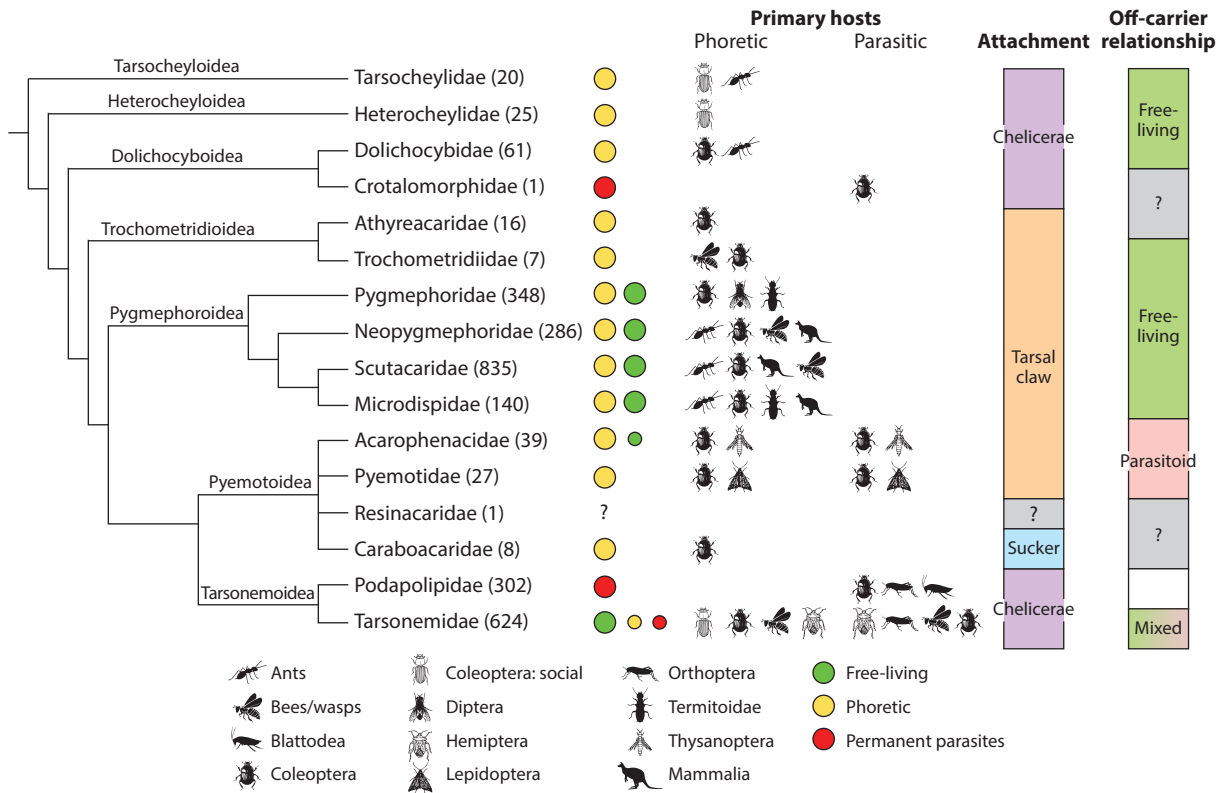


Figure 2

Phylogeny of Heterostigmata (84): species diversity (updated from Reference 148), primary host relationships (smaller circles indicate less representation), primary carriers or hosts (up to four, from most to least important), attachment method of phoretics, and primary off-carrier relationship.

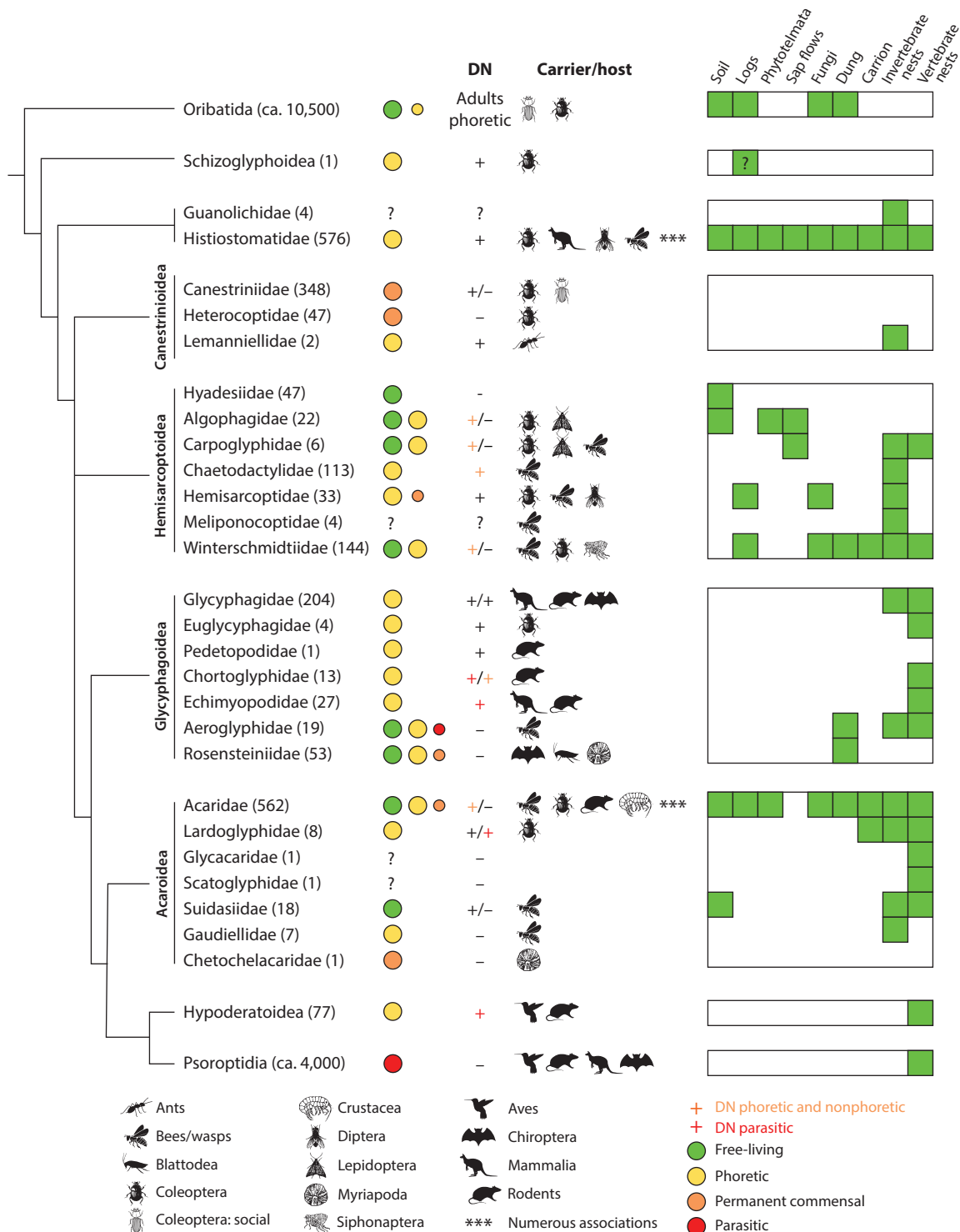
However, in later branches, egg parasitoidism is prevalent, especially in the Pyemotoidea, where adult females ride a host and attack eggs as they are laid.

Early derivative taxa attach to carriers with their chelicerae (Tarsocheilidae, Heterocheilidae, Dolichocyboidea), as do the derived Tarsonemoidea (**Figure 2**). Permanent parasitism arises only in Tarsonemoidea (58, 84), notably the Podapolipidae, a radiation found primarily on beetles and orthopteroids (118, 131). Attachment to carriers via chelicerae may thus be a preadaptation to parasitism. Within most Trochometridioidea, Pygmephoroidae, and Pyemotoidea, attachment occurs via a claw-like structure at the tip of the tarsus (often a phoretomorph) that clasps the setae of the carrier. Notable variants are the bizarre Caraboacaridae, in which females attach via chelicerae and coxal suckers (59), and *Paracarophenax* (Acarophenacidae), which secrete a glue-like plug that they pinch for attachment (143).

The Heterostigmata present a shift from phoresy back to free-living lifestyles, a surprising theme also found in the other major groups exhibiting phoresy. The Tarsonemidae have made the shift most spectacularly, with numerous nonphoretic fungivorous and phytophagous taxa among phoretic lineages (84). Of the phoretic taxa, the best known are those within the nests of scolytine beetles that spread pathogenic bluestain fungi in sporothecae (48). However, some tarsonemids have more intimate or curious symbioses with their carriers. *Iponemus* are egg parasitoids of their scolytine beetle carriers (83), several tarsonemellines cause galls in figs and are transported on

Phoretomorph:
a heteromorphic phoretic stage with a morphology differing from nonphoretic forms of the same stage

Sporothecae:
special pockets for transporting ascospores



(Caption appears on following page)

Figure 3 (Figure appears on preceding page)

Phylogeny of Astigmata (105): species diversity (updated from Reference 123), primary host relationship (smaller circles indicate less representation), presence of deutonymphs (DN) [optional (*orange* +) or parasitic (*red* +)], primary carriers (up to four, from most to least important), and natal habitats for free-living and phoretic taxa.

fig wasps (56), and the wind-dispersed phytophagous pest *Polyphagotarsonemus latus* is phoretic on whiteflies (112) (**Figure 1c**).

2.1.3. The Astigmata. The Astigmata present at least three remarkable shifts: first, from a probable edaphic free-living oribatid ancestor to a mite with a highly specialized deutonymph specifically adapted for phoresy; second, to permanent parasites of vertebrates (Psoroptidia); and third, from vertebrate parasites to nest inhabitants (Pyroglyphoidea, the house-dust mites) (26, 65, 66). The phylogenetic relationships of lower Astigmata, which are primarily associated with insects, are poorly resolved. In this review, we use previous morphological hypotheses (101) but acknowledge that more inclusive molecular studies will soon provide better resolution (26, 66).

Unlike Heterostigmata, Astigmata have one primary attachment method via a ventral plate of suckers (**Figure 1a,b**), although some groups have further modifications for clasping onto mammal hair (106). Purported early taxa (Schizoglyphoidea, Histiotomatoidea, Canestrinioidea, Hemisarcopitoidea) tend to be found on insects that utilize edaphic ephemeral habitats, but this pattern is less striking than in the Heterostigmata and Mesostigmata. Nevertheless, mammal and bird associates, with the natal habitat of nests, are primarily found in the Glycyphagoidea, Acaroidea, and Hypoderatoidea (**Figure 3**). In contrast to the Mesostigmata and Heterostigmata, beetle associations are less common than associations with bees and wasps, and few Astigmata are intimately associated with ants. However, the most diverse families (Acaridae, Histiotomatidae) often attach, probably in nonspecific relationships, to various insects as well as terrestrial vertebrates.

Some Astigmata show intraspecific life stage plasticity in the deutonymph, which is skipped or takes a phoretic or sedentary form (71, 72, 104). Skipping the deutonymph allows for rapid population growth and thus resource exploitation, while the sedentary and phoretic forms provide security in time and space, respectively. Such plasticity is expected more often in cyclical or longer-lasting habitats with harsh periods or irregular resource replenishment (4, 37), characteristic of many habitats utilized by phoretic Astigmata.

2.1.4. Other independent derivations. Phoresy has evolved at least 10 times in the remaining Acariformes. The Oribatida are infrequently phoretic, but several wood-inhabiting species have adults that are phoretic on saproxylic beetles. Passalid beetles are the most common carriers, particularly of the Mesoplophoridae, which clamp onto setae with their folding, jackknife bodies (100). More passive phoresy occurs in other families, such as Oppiidae, Scheloribatidae, and Haplozetidae (34, 70, 122). Curiously, several Oppiidae are phoretic on dung beetles, an atypical habitat for oribatids (33), and myrmecophilous species must somehow move between ant nests (54).

In the Trombidiformes, phoretic relationships occur in some Cheyletidae (17), Ereyneidae (35) (**Figure 1e**), and Iolinidae (140). Furthermore, other taxa form loose or poorly studied relationships with insects, such as Tydeidae on moths (140), Stigmaeidae on sandflies (149), and Dasythyreidae on click beetles (142). The phytophagous Eriophyoidea are wind dispersed, although pollinators may transport some species between host plants (41); more extraordinary are phoretic *Aceria pallida* that accompany their psyllid carrier not directly between host plants but to overwintering sites (87).

Life stage plasticity:
the ability to vary
developmental
patterns according to
environmental
conditions

2.2. Parasitiformes

In contrast to the Acariformes, the earliest parasitiform fossil is Cretaceous (57). An earlier origin is expected, yet the lack of fossils, in tandem with a strong Gondwanan pattern for one of the earliest branches—the Holothyrida + Ixodida—gives credence to a late Cretaceous or early Cenozoic diversification (9). Whenever their origin, phoretic associations with insects are present within the earliest branches of Mesostigmata: Trigynaspida, Microgynioidea, Sejoidea, and Uropodina (**Figure 4**).

The Mesostigmata show three distinctive types of phoretic relationships (**Figure 4**). The first and earliest type comprises the Trigynaspida, which are phoretic as adults on insects, especially ants and wood-inhabiting beetles (62) (**Figure 1f**). The exceptions are several free-living Cercomegistoidea that may represent prephoretic ancestors. Many trigynaspid mites live on their hosts even while their hosts are not dispersing and are thus in another nonphoretic symbiosis. Where they have been studied, the Celaenopsoidea, Cercomegistoidea, Fedrizzioidea, and Megisthanoidea are predators and scavengers and are thus commensal (20, 63, 128, 139); the other superfamilies are kleptoparasitic on carabid beetles or ants (40). Trigynaspids tend to live in long-lasting habitats, probably due to their relatively long life cycles of at least one month (63, 128, 139). They usually have no special adaptations for attachment and move freely over their hosts. Parasitic adults, such as the Paramegistidae (27), have evolved multiple times.

The second type involves several seemingly independent derivations of phoretic deutonymphs. The first of these are the Sejina and Uropodina, currently seen as two independent lineages that independently developed deutonymphs that attach via an anal pedicel (69) (**Figure 1g**). As in trigynaspids, wood-inhabiting beetles are common hosts (60), as are ants for Uropodina (120). Both groups prefer long-lived habitats such as rotting wood and nests. The second group is a radiation in the otherwise free-living Parasitidae, the Parasitinae, which are phoretic as relatively unspecialized deutonymphs (**Figure 1e,b**). Due to the relatively fast life cycles of parasitines, their natal habitats are often carrion, dung, and nests, and their carriers tend to be insects living in these habitats (74, 126). The Rhodacaroidea continue the deutonymphal pattern into the Gamasina: Some Ologamasidae are phoretic on mammals and inhabit their nests or are phoretic on carabid beetles; Halolaelapidae are phoretic on amphipods, beetles, and flies in wet habitats; and a large radiation of Digamasellidae are phoretic primarily on saproxylic beetles (85).

The third type encompasses four highly diverse groups of Mesostigmata: Ascoidea, Dermanyssoidea, Eviphidoidea, and Phytoseioidea. These mites are phoretic as adults and tend to have fast life cycles (approximately one week) (142). As such, these Mesostigmata are found primarily in carrion, dung, flowers, or other resources that quickly degrade (75, 85). However, this does not preclude them from exploiting longer-lasting habitats, especially the nests of insects (31, 107).

3. ADAPTATIONS FOR PHORESY: INNOVATION AND CONVERGENCE

Phoresy evolved a surprising number of times and in relationships with almost all winged insect orders, but primarily with beetles and nest-making Hymenoptera (**Figures 2–4**). The natal habitats of phoretic species are diverse, but phoresy appears to have initially evolved when a free-living edaphic species with a slow life cycle exploited the habitat of rotting logs. This habitat is among the longest lasting of all patchy environments (91). We hypothesize that rotten logs present an evolutionarily solvable problem: That is, dispersal between logs on foot could still be successful, but phoresy is more successful and therefore selected for. The three major phoretic groups, as well as Oribatida, share this ancestral habitat. The Parasitengona are an exception, rarely parasitizing log-inhabiting or nesting insects (38).

Haplodiploid:

a sex-determination system in which haploid males develop from unfertilized eggs, and diploid females develop from fertilized eggs

Phoretic parasite:

species that is parasitic in one life stage while also being dispersed by its host

After rotting logs, phoretic species exploited all manner of habitats, including those that persist only for a matter of days or weeks. The key innovation seems to be faster life cycles (104). A mite's life cycle must be shorter than the duration of the resource. Thus, mites with slow life cycles, such as trigynaspids (128, 139) and oribatids (101), occupy long-lasting habitats. In contrast, the faster life cycles of higher Mesostigmata, Heterostigmata, and Astigmata—as little as a week (e.g., 78, 116, 126)—allow exploitation of short-lived resources such as dung, carrion, flowers, and sporocarps (104). These higher taxa also utilize long-lasting habitats, such as tree holes (37) and sap flows (44), although they tend to have slower life cycles.

The phoretic life stage is usually an adult or deutonymph. Phoretic adults are often females (88, 141), but both sexes are phoretic in species that have nonphoretic periods on their host (e.g., Trigynaspida; 130) or where males seek additional mates in new habitats (e.g., 42, 119). Phoresy by adults provides obvious advantages: Mated adult females or those with haplodiploid genetic systems (102) can establish colonies alone and exploit resources instantaneously. More significantly, their progeny can feast on ephemeral prey that also exploit the resource (e.g., nematodes, eggs, early-instar larvae). Thus, phoresy by adults dominates in species that exploit short-lived resources, whether it be the resources that the habitat presents (e.g., small prey, nectar) or the host's progeny (i.e., host eggs or provisions). Why do some species disperse as deutonymphs? We think that it is significant that the most morphologically specialized phoretic mites are deutonymphs. Immature mites do not need to find mates and bear young, and the specialized deutonymphs of Astigmata and Uropodina do not feed, allowing investment in adaptations for phoresy. The existence of a phoretic morph resistant to adverse conditions also permits some environmental resilience and life-cycle synchronization, which are ideal for life in a long-lived cyclic habitat. Furthermore, deutonymphs are smaller than adults and, in bisexual species, must disperse in groups to increase mating opportunity. If carrying mites incurs a cost to the insect, then perhaps having smaller nymphs reduces these impacts.

Unusually, protonymphs are never the phoretic stage, and phoretic larvae exist only as the phoretic parasites of Parasitengona. We hypothesize that enough resources must be accumulated to invest in a specialized and resilient life stage, as in the Astigmata, or the preadult stage, as in the Sejina, Uropodina, Parasitidae, and basal Gamasina.

4. PHORESIS AND THE DIVERSIFICATION OF MITES

Phoresy allows mites to exploit a plethora of resources that are otherwise unobtainable, permitting numerous radiations throughout the three major lineages that have evolved phoresy. Thus, phoretic mites contribute significantly to the diversity of the Acari. However, free-living and parasitic mites outnumber phoretic mites significantly: Almost all Oribatida are free living (101), and most Trombidiformes and Mesostigmata are free-living or permanent parasites (**Figures 2–4**). Therefore, phoresy itself has not promoted unusual rates of speciation. Indeed, the greater dispersal ability may promote panmixis in populations, reducing fragmentation as experienced by soil-dwelling free-living mites. In permanent parasites, host specificity and on-host microhabitat specificity likely promote speciation. Phoretic mites could experience similar pressures, but probably only in nests and environments where the natal habitat requires species-specific adaptations for tracking the life cycle of hosts, and where options for carriers are limited.

5. INTERACTIONS BETWEEN MITES AND THEIR CARRIERS

Phoresy is a temporary relationship that is frequently regarded as commensal. As mites rely on their carrier for safe transit, the smooth, flattened bodies of many phoretic mites may have evolved to minimize impact on their carrier. However, numerous phoretic mites have bodies with little

modification, and those of Uropodina attach via stalks, thus nullifying aerodynamic benefits to their carrier. Occasionally, mite loads hinder the movement of their carrier (e.g., 64, 89), and some mites feed on host tissues in transit (e.g., 51, 115) or prey on phoretic nematodes (76). Despite these few examples of negative impacts or feeding, the mite is still phoretic: It is a single life stage moving between habitats on a carrier.

Phoretic mites rely on their carrier for transport, and in most cases, the natal habitat for mites is the same as their carrier. Therefore, we expect insignificant or even beneficial symbioses (43, 107, 145), particularly cleaning and protective mutualisms (e.g., 13, 108), in the shared natal habitat. However, numerous examples of negative symbioses exist. For instance, the heterostigmatic families Pyemotidae and Acarophenacidae contain parasitoids of the eggs of their carriers (32, 98), *Chaetodactylus* kills the young of mason bees (94), *Trochometridium* kills the larva of various bees and then feeds on the resulting fungal growth (from spores carried by the mites) (24), and *Macrocheles muscaedomesticae* eats the eggs and first-instar larvae of its muscid fly carrier (39). In this section, we discuss four well-studied systems: the mite–fungus community of scolytine bark beetles, the hummingbird–flower mites, *Poecilobirus* on carrion beetles, and mites in the acarinaria of bees.

The community of mites living with bark beetles is a complex system of several species of mites, fungi, and beetles (47, 48, 113). Most work focuses on southern pine beetle (*Dendroctonus frontalis*) and its phoretic mites, particularly *Tarsonemus*. Beetles and *Tarsonemus* both actively disperse wood-pathogenic fungi upon which the beetle larvae and mites rely. However, mites also actively transport the beetle-antagonistic bluestain fungus, which can outcompete beetle-mutualistic fungi. Thus, the *Tarsonemus*–beetle relationship has mutualistic benefits, by supplementing transmission of beneficial fungi, that are weak at best; this relationship instead tends to be indirectly antagonistic when *Tarsonemus* transmits bluestain fungi (45, 46). Indeed, evidence for mite–beetle mutualisms is weak and is only inferred from the vast diversity of mites with bark beetles, some of which presumably prey on *Tarsonemus*; however, others are predatory of their own carriers (96).

Several groups of Mesostigmata have colonized flower–pollinator systems, where they consume nectar and pollen and are phoretic on floral visitors (19, 42, 95, 127). The best known of these mites are the hummingbird–flower mites, which dash up the bills of hummingbirds and hide in their nares until the hummingbird visits the next suitable flower (19, 22). Unlike the other systems, the mites do not share the natal habitat directly; however, they compete with hummingbirds for the same nectar, removing up to half of the resource (79) and potentially altering foraging behavior (80). Thus, hummingbird–flower mites rob resources from their carrier while providing no benefit in return.

The mites of both carrion beetles and eumenine wasps are examples of protective mutualisms, where mites attack competing flies and parasitic wasps, respectively. However, this harmonious relationship has a sinister side. In both cases, the relationship is context dependent, shifting between mutualistic and antagonistic according to biotic and/or physical conditions.

Carrion beetles (*Nicrophorus*) compete with calliphorid flies for the corpses of small mammals, but only carrion beetles regularly carry parasitoid mites (*Poecilobirus* spp.; **Figure 1b**). These mites feed on the eggs of calliphorid flies and are thus beneficial to beetles, but the mites also attack the eggs of their hosts (12, 15) and compete for carrion (99, 124, 137). The mites are thus in a conditional relationship with their hosts, being disadvantageous in the absence of blowflies but often advantageous when blowflies are present (137, 145).

Several bees and wasps have pits or pockets (acarinaria) in which they carry mites. Given that these structures seemingly encourage phoretic mites, surely these mites should be beneficial. However, numerous mites riding in acarinaria feed on their carrier's progeny or provisions (23; <http://idtools.org/id/mites/beemites/index.php>). Thus, instead of transporting beneficial mites, acarinaria could instead be a structure for skewing mite loads as social bees depart their natal

Context dependent:

describes a symbiosis that is conditional, changing its nature according to certain biotic or physical factors

nest (67). Unfortunately, we know little about the impact of most of these mites; one of the best-studied relationships is that of the eumenine wasp *Allodynerus delphinalis* and its mite *Ensliniella parasitica* (108–110). These mites feed on the hemolymph of juvenile wasps and, at high densities, cause mortality of their carrier (110). However, at lower densities, the mites have little impact and, when the parasitoid wasp *Melittobia acasta* is present, defend juvenile wasps by attacking and sometimes killing the parasitoid (108).

The above examples show how assumptions of relationships based on feeding or a limited number of observations can be misleading. In each example, the multitrophic interactions are more complex than was first thought: Few, if any, phoretic mites exist in a single phoretic-carrier system, and some are extraordinarily complex. For example, beetles and sunbirds transport four phoretic mite species, with the mites themselves transporting fungal spores, to protea flowers, where only some mite species utilize the fungi for food (138). The interactions among players in these multipartite symbioses are difficult to disentangle, but phoretic mites tend to build exploitative relationships with their carriers that are only mutualistic under certain conditions. Nevertheless, stable commensal or mutualistic relationships may occur; however, these have been little studied, and we encourage work on the impact of more mundane phoretics on their carriers.

6. EVOLUTIONARY TRANSITIONS FROM PHORESY

Phoresy is a logical intermediate life history because it requires elements of parasitism (host finding and attachment) but no adaptations for feeding, breeding, and transfer between hosts (3, 5, 49, 132). Phylogenetic evidence generally supports this pivotal position, as lineages of permanent symbionts arise consistently within or as sister to lineages exhibiting phoresy (**Figures 2–4**). Significant exceptions are deep branches, most notably the ticks and Parasitengona, which have no known phoretic intermediary (9, 146).

Two pathways to parasitism are possible: via the phoretic symbiosis itself or via the off-host symbiosis. The former involves the phoretic stage first becoming a phoretic parasite, while the latter involves other stages shifting to parasitism, or another permanent symbiosis, while maintaining a phoretic stage. The relative importance of each pathway is difficult to disentangle, and we suspect that both pathways have played a significant role.

Examples of phoretic mites sometimes feeding from their hosts, such as *Macrocheles* (90, 114, 115) and *Hemisarcopetes cooremani* (51), are purported transitional relationships leading to parasitism. However, we see these relationships as a step toward obligate phoretic parasitism; i.e., feeding on the carrier is required for further development. For the next step, phoretic parasites must abandon their ancestral feeding habits and shift to living and breeding on their hosts. However, examples of this transition are elusive. Water mites with parasitic nymphs and adults do not parasitize their chironomid carrier but instead shift to mollusks (30) or sponges (25). In the Glycyphagoidea (Astigmata), the specialized deutonymphs were originally insect phoretics, but a putative transformation series from vertebrate phoretics to endofollicular phoretics and finally to endofollicular phoretic parasites occurred. None, however, have abandoned the natal habitat of the nest. Similarly, the Hypoderatidae are regressive deutonymphal parasites within the tissues of birds and rodents, but, where they are known, active stages develop in nests. Therefore, phoretic parasites with highly modified life histories and morphologies may not transition easily to permanent parasitism, but, as in the example of the Hypoderatidae, the probable sister to the Psoroptidia (103), when they do so it may allow an extraordinary radiation.

A shift toward parasitism by nonphoretic stages first requires a more intimate off-host relationship, of which a nest is the most intimate of all. Significant lineages of bird and mammal parasites may have followed this pathway, such as the Dermanyssoidea (117), Cheyletoidea (16), and Psoroptidia (6). The same pathway seems likely for some permanent symbionts of insects

found on nesting beetles (e.g., Diarthrophallidae on Passalidae) and social bees (e.g., Varroidae on Apidae) and Uropodina with unmodified deutonymphs in ant colonies (77). However, some permanent symbionts of insects have no links with nests, such as the Otopheidomeninae (moths; 86) and Podapolipidae (chrysomelid beetles; 129) or have ambiguous ancestral host relationships (e.g., Canestriniidae, possibly on carabids; 105). Thus, these lineages may have arisen through phoretic parasitism by adults, coupled with selection to remain on hosts that did not utilize concentrated resources, thus making new hosts difficult to locate.

Phoresy is readily reversible, with free-living taxa arising frequently within otherwise phoretic lineages (**Figures 2–4**). In Heterostigmata, each family of Pygmephorodea reverted to free living (8), as have Tarsonemidae and possibly the Acarophenacidae. Surprisingly, several Astigmata and Uropodina also reverted to free living, despite their heteromorphic phoretic deutonymphs. Most Astigmata accomplished this reversion by skipping the deutonymph, which often shows plasticity in its expression (71). Similarly, some Uropodina have phoretic and nonphoretic deutonymphs (2), and adult phoretomorphs occur in some Heterostigmata (97) and Mesostigmata (10). The optional expression of these stages suggests that they could be lost easily. Phoretic parasitism is also lost frequently in the Parasitengona, primarily within water mites (135) but also in terrestrial groups (147). Thus, phoresy has evolved frequently as a means of exploiting patchy resources, but it seems to be just as easily lost, representing reversions to free-living lifestyles where food is presumably harder to find but more evenly distributed.

SUMMARY POINTS

1. Phoresy is a temporary symbiosis where a larger animal transports a single life stage of a smaller animal.
2. Feeding may occur on the carrier and is obligate in the larval parasites of Parasitengona, deutonymphal vertebrate parasites of the Astigmata, and some kleptoparasitic and parasitic Mesostigmata.
3. Rotting logs are an ancestral habitat, with faster life cycles in derived taxa allowing exploitation of shorter-lived resources.
4. Phoresy is usually part of another symbiosis with the carrier, which can be a complex, context-dependent relationship that nevertheless tends to be exploitative of the carrier's progeny or its resources.
5. Phoresy has evolved multiple times—although just once in Astigmata and perhaps Heterostigmata—but is also lost frequently.
6. Permanent parasitism has evolved from numerous phoretic lineages but rarely, if ever, from lineages with parasitic phoretic stages.

DISCLOSURE STATEMENT

The authors are not aware of any affiliations, memberships, funding, or financial holdings that might be perceived as affecting the objectivity of this review.

ACKNOWLEDGMENTS

We are grateful for the constructive discussion and comments from Drs. Frederic Beaulieu (Canadian National Collection of Insects, Arachnids and Nematodes, Ottawa) and Helen Nahrung

(University of the Sunshine Coast, Sippy Downs) and the superb photos supplied by the iNaturalist community.

LITERATURE CITED

1. Athias-Binche F. 1984. La phorésie chez les acariens uropodides (Anactinotriches), une stratégie écologique originale. *Acta Ecol. Ecol. Gen.* 5:119–33
2. Athias-Binche F. 1984. Phoresy in Uropodina (Anactinotrichida), occurrence, demographic involvements and ecological significance. In *Acarology*, Vol. 1, ed. DA Griffiths, CE Bowman, pp. 276–85. Chichester, UK: Ellis Horwood
3. Athias-Binche F. 1994. *La Phorésie chez les acariens. Aspects adaptatifs et évolutifs*. Perpignan, France: Éd. Castillet
4. Athias-Binche F. 1995. Phenotypic plasticity, polymorphisms in variable environments and some evolutionary consequences in phoretic mites (Acarina): a review. *Ecologie* 26:225–41
5. Athias-Binche F, Morand S. 1993. From phoresy to parasitism: the example of mites and nematodes. *Res. Rev. Parasitol.* 53:73–79
6. Atyeo WT, Gaud J. 1979. Feather mites and their hosts. In *Recent Advances in Acarology*, Vol. II, ed. JG Rodriguez, pp. 355–61. New York: Academic
7. Bartlow AW, Agosta SJ. 2021. Phoresy in animals: review and synthesis of a common but understudied mode of dispersal. *Biol. Rev.* 96:223–46
8. Baumann J. 2018. Tiny mites on a great journey—a review on scutacarid mites as phoronts and inquilines (Heterostigmatina, Pygmephoroidae, Scutacaridae). *Acarologia* 58:192–251
9. Beati L, Klompen H. 2019. Phylogeography of ticks (Acari: Ixodida). *Annu. Rev. Entomol.* 64:379–97
10. Beaulieu F, Déchêne AD, Walter DE. 2008. Phase morphs and phoresy: new species of *Antennoseius* (*Vitzthumia*) mites (Acari: Mesostigmata: Ascidae) associated with pyrophilous carabids (Carabidae: *Sericoda* spp.) in Alberta, Canada. *Zootaxa* 1961:37–57
11. Beaulieu F, Dowling APG, Klompen H, de Moraes GJ, Walter DE. 2011. Superorder Parasitiformes Reuter, 1909. *Zootaxa* 3148:123–28
12. Beninger CW. 1993. Egg predation by *Poecilochirus carabi* (Mesostigmata: Parasitidae) and its effect on reproduction of *Nicrophorus vespilloides* (Coleoptera: Silphidae). *Econ. Entomol.* 22:766–69
13. Biani NB, Mueller UG, Wcislo WT. 2009. Cleaner mites: sanitary mutualism in the miniature ecosystem of Neotropical bee nests. *Am. Nat.* 173:841–47
14. Binns ES. 1982. Phoresy as migration—some functional aspects of phoresy in mites. *Biol. Rev.* 57:571–620
15. Blackman SW. 1997. Experimental evidence that the mite *Poecilochirus davydovae* (Mesostigmata: Parasitidae) eats the eggs of its beetle host. *J. Zool. Lond.* 242:63–67
16. Bochkov AV. 2004. Mites of the family Cheyletidae (Acari: Prostigmata): phylogeny, distribution, evolution and analysis of parasite-host relationships. *Parazitologiya* 38:122–38
17. Bochkov AV, Klimov PB. 2005. Three new species of the predaceous Cheyletidae (Acari: Prostigmata) phoretic on insects. *Acarina* 13:15–22
18. Bohonak AJ, Smith BP, Thornton M. 2004. Distributional, morphological and generic consequences of dispersal for temporary pond water mites. *Freshw. Biol.* 49:170–80
19. Britto EPJ, Finotti AS, de Moraes GJ. 2015. Diversity and population dynamics of Ascidae, Blattisociidae and Melicharidae (Acari: Mesostigmata) in tropical flowers in Brazil. *Exp. Appl. Acarol.* 66:203–17
20. Butler L, Hunter PE. 1968. Redescription of *Megisthanus floridanus* with observations on its biology (Acarina: Megisthanidae). *Fla. Entomol.* 51:187–97
21. Camerik AM. 2010. Phoresy revisited. In *Trends in Acarology*, ed. M Sabelis, J Bruin, pp. 333–36. Berlin: Springer
22. Colwell RK. 1973. Competition and coexistence in a simple tropical community. *Am. Nat.* 107:737–60
23. Cowan DP. 1984. Life history and male dimorphism in the mite *Kennethiella trisetosa* (Acarina: Winterschmidtidae), and its symbiotic relationship with the wasp *Ancistrocerus antilope* (Hymenoptera: Eumenidae). *Ann. Entomol. Soc. Am.* 77:725–32

24. Cross EA, Bohart GE. 1978. Some observations of the habits and distribution of *Trochometridium* Cross, 1965 (Acarina: Pyemotidae). *Acarologia* 20:286–93
25. Crowell RM, Davids C. 1979. Systematics of *Unionicola laurentiana*, n.sp., and *U. nearctica*, n.sp., sponge-associated Hydracarina (Parasitengona: Unionicolidae) from North America. *Ohio J. Sci.* 79:178–86
26. Dabert M, Witalinski W, Kazmierski A, Olszanowski Z, Dabert J. 2010. Molecular phylogeny of acariform mites (Acari, Arachnida): strong conflict between phylogenetic signal and long-branch attraction artifacts. *Mol. Phylogenet. Evol.* 56:222–41
27. Derne BT, Hutchinson MN, Weinstein P, Gardner MG, Halliday B. 2018. Parasite in peril? A new species of mite in the genus *Ophiomegistus* Banks (Parasitiformes: Paramegistidae) on an endangered host, the pygmy bluetongue lizard *Tiliqua adelaidensis* (Peters) (Squamata: Scincidae). *Aust. Ecol.* 44:420–32
28. Dowling APG, O'Connor BM. 2010. Phylogenetic relationships within the suborder Dermanyssina (Acari: Parasitiformes) and a test of dermanyssoid monophyly. *Int. J. Acarol.* 36:299–312
29. Dunlop JA, Frahnert K, Mąkol J. 2018. A giant mite in Cretaceous Burmese amber. *Foss. Rec.* 21:285–90
30. Edwards DD, Dimock RV Jr. 1995. Specificity of the host recognition behaviour of larval *Unionicola* (Acari: Unionicolidae): the effects of larval ontogeny and early larval experience. *Anim. Behav.* 50:343–52
31. Eickwort GC. 1990. Associations of mites with social insects. *Annu. Rev. Entomol.* 35:469–88
32. Elbadry EA, Tawfik MSF. 1966. Life cycle of the mite *Adactylidium* sp. (Acarina: Pyemotidae), a predator of thrips eggs in the United Arab Republic. *Ann. Entomol. Soc. Am.* 59:458–61
33. Ermilov SG, Frolov AV. 2019. *Ramusella* (*Dosangoppia*) *bochkovi* (Acari, Oribatida, Oppiidae), a new subgenus and species of oribatid mites phoretic on *Ceratophyus polyceros* (Pallas, 1771) (Coleoptera, Geotrupidae) from Russia. *Syst. Appl. Acarol.* 23:209–21
34. Ermilov SG, Frolov AV. 2021. New data on oribatid mites (Acari, Oribatida) phoretic on passalid beetles (Coleoptera, Passalidae) from the Afrotropical and Oriental regions, with descriptions of three new species from Congo, Gabon and Ghana. *Syst. Appl. Acarol.* 26:769–87
35. Fain A, Camerik AM. 1994. Notes on the mites of the genus *Ereynetes* Berlese (Acari: Ereynetinae), with description of five new species from South Africa. *Bull. Inst. R. Sci. Nat. Belg.* 64:145–64
36. Farish DJ, Axtell RC. 1971. Phoresy redefined and examined in *Macrocheles muscaedomesticae* (Acarina: Macrochelidae). *Acarologia* 13:16–29
37. Fashing NJ. 2010. Life history and biology of *Hormosianoetus mallotae* (Fashing) (Histiostomatidae: Astigmata), an obligatory inhabitant of water-filled treeholes. *Int. J. Acarol.* 36:189–98
38. Felska M, Wohltmann A, Makol J. 2018. A synopsis of host-parasite associations between Trombidioidea (Trombidiformes: Prostigmata, Parasitengona) and arthropod hosts. *Syst. Appl. Acarol.* 23:1375–479
39. Filipponi A. 1955. Sulla natura dell'associazione tra *Macrocheles muscaedomesticae* e *Musca domestica*. *Riv. Parasitol.* 16:83–102
40. Franks NR, Healey KJ, Byrom L. 1991. Studies on the relationship between the ant ectoparasite *Antennophorus grandis* (Acarina: Antennophoridae) and its host *Lasius flavus* (Hymenoptera: Formicidae). *J. Zool. Lond.* 225:59–70
41. Galvão AS, Melo JWS, Monteiro VB, Lima DB, de Moraes GJ, Gondim MGC Jr. 2012. Dispersal strategies of *Aceria guerreronis* (Acari: Eriophyidae), a coconut pest. *Exp. Appl. Acarol.* 57:1–13
42. Guerra TJ, Romero GQ, Costa JC, Lofego AC, Benson WW. 2012. Phoretic dispersal on bumblebees by bromeliad flower mites (Mesostigmata, Melicharidae). *Insect Soc.* 59:11–16
43. Halliday RB. 2019. The enemy of my parasite is my friend: the possible role of predatory mites as biological control agents of pest beetles in soil. *Int. J. Acarol.* 45:189–96
44. Hayashi K, Ichikawa T, Yasui Y. 2010. Life history of the newly discovered Japanese tree sap mite, *Hericia sanukiensis* (Acari, Astigmata, Algophagidae). *Exp. Appl. Acarol.* 50:35–49
45. Hofstetter RW, Cronin J, Klepzig KD, Moser JC, Ayres MP. 2006. Antagonisms, mutualisms and commensalisms affect outbreak dynamics of the southern pine beetle. *Oecologia* 147:679–91
46. Hofstetter RW, Dempsey TD, Klepzig KD, Ayres MP. 2007. Temperature-dependent effects on mutualistic and phoretic associations. *Community Ecol.* 8:47–56
47. Hofstetter RW, Dinkins-Bookwalter J, Davis TS, Klepzig KD. 2015. Symbiotic associations of bark beetles. In *Bark Beetles: Biology and Ecology of Native and Invasive Species*, ed. FE Vega, RW Hofstetter, pp. 209–45. New York: Academic

48. Hofstetter RW, Moser JC. 2014. The role of mites in insect-fungus associations. *Annu. Rev. Entomol.* 59:537–57
49. Houck MA. 1994. Adaptation and transition into parasitism from commensalism: a phoretic model. See Ref. 50, pp. 252–81. Berlin: Springer
50. Houck MA, ed. 1994. *Mites: Ecological and Evolutionary Analyses of Life-History Patterns*. Berlin: Springer
51. Houck MA, Cohen AC. 1995. The potential role of phoresy in the evolution of parasitism: radiolabelling (tritium) evidence from an astigmatid mite. *Exp. Appl. Acarol.* 19:677–94
52. Houck MA, O'Connor BM. 1991. Ecological and evolutionary significance of phoresy in the Astigmata. *Annu. Rev. Entomol.* 36:611–36
53. Hunter PE, Rosario RMT. 1988. Associations of Mesostigmata with other arthropods. *Annu. Rev. Entomol.* 33:393–417
54. Ito F. 2013. Evaluation of the benefits of a myrmecophilous oribatid mite, *Aribates javensis*, to a myrmecine ant, *Myrmecina* sp. *Exp. Appl. Acarol.* 61:79–85
55. Janet C. 1897. *Etudes sur les Fourmis, les Guêpes et les Abeilles. Note 14, Rapports des Animaux Myrmécophiles avec les Fourmis*. Limoges, Fr.: VH Ducourtieux
56. Jauharlina J, Lindquist EE, Quinnell RJ, Robertson HG, Compton SG. 2012. Fig wasps as vectors of mites and nematodes. *Afr. Entomol.* 20:101–10
57. Joharchi O, Vorontsov D, Walter DE. 2021. Oldest determined record of a mesostigmatic mite (Acari: Mesostigmata: Sejidae) in Cretaceous Burmese amber. *Acarologia* 61:641–49
58. Kaliszewski M, Athias-Binche F, Lindquist EE. 1995. Parasitism and parasitoidism in Tarsonemina (Acari: Heterostigmata) and evolutionary considerations. *Adv. Parasitol.* 35:335–67
59. Katlav A, Hajiqaanbar H, Talebi AA. 2015. A new genus and species of mites of the family Carabacaridae (Acari: Heterostigmata) associated with *Clivina ypsilon* (Coleoptera: Carabidae) with notes on distribution and host range of the family. *Can. Entomol.* 147:370–80
60. Kethley JB. 1983. Modifications of the deutonymph of *Uropodella lacinata* Berlese, 1888, for phoretic dispersal (Acari: Parasitiformes). *J. Ga. Entomol. Soc.* 18:151–55
61. Khaustov AA, Poinar G Jr. 2011. *Protoresinacarus brevipedis* gen. n., sp. n. from early Cretaceous Burmese amber: the first fossil record of mites of the family Resinacaridae (Acari: Heterostigmata: Pyemotoidea). *Hist. Biol.* 23:219–22
62. Kim CM. 2004. Trigynaspida (Acari: Mesostigmata): new diagnosis, classification and phylogeny. *Acarologia* 44:157–94
63. Kinn DN. 1971. *The Life Cycle and Behaviour of Cercoleipus coelonotus (Acarina: Mesostigmata) Including a Survey of Phoretic Mite Associates of California Scolytidae*. Univ. Calif. Publ. Entomol. 65. Berkeley, CA: Univ. Calif. Press
64. Kinn DN, Witcosky JJ. 1978. Variation in southern pine beetle attack height associated with phoretic uropodid mites. *Can. Entomol.* 110:249–51
65. Klimov PB, O'Connor B. 2013. Is permanent parasitism reversible? Critical evidence from early evolution of house dust mites. *Syst. Biol.* 62:411–23
66. Klimov PB, O'Connor B, Chetverikov PE, Bolton SJ, Pepato AR, et al. 2018. Comprehensive phylogeny of acariform mites (Acariformes) provides insights on the origin of the four-legged mites (Eriophyoidea), a long branch. *Mol. Phylogenet. Evol.* 119:105–17
67. Klimov PB, Vinson SB, O'Connor BM. 2007. Acarinaria in associations of apid bees (Hymenoptera) and chaetodactylid mites (Acari). *Invertebr. Syst.* 21:109–36
68. Klimov PB, Vorontsov DD, Azar D, Sidorchuk EA, Braig HR, et al. 2021. A transitional fossil mite (Astigmata: Levantoglyphidae fam. n.) from the early Cretaceous suggests gradual evolution of phoresy related metamorphosis. *Sci. Rep.* 11:15113
69. Klompen H, Lekveishvili M, Black WC IV. 2007. Phylogeny of parasitiform mites (Acari) based on rRNA. *Mol. Phylogenet. Evol.* 43:936–51
70. Knee W. 2017. A new *Paraleius* species (Acari, Oribatida, Scheloribatidae) associated with bark beetles (Curculionidae, Scolytinae) in Canada. *ZooKeys* 667:51–65
71. Knülle W. 1987. Genetic variability and ecological adaptability of hypopus formation in a stored product mite. *Exp. Appl. Acarol.* 3:21–32

72. Knülle W. 2003. Interaction between genetic and inductive factors controlling the expression of dispersal and dormancy morphs in dimorphic astigmatic mites. *Evolution* 57:828–38
73. Konikiewicz M, Małol J. 2018. Insight into the fossil fauna of terrestrial Parasitengona mites (Trombidiformes: Prostigmata)—the first representatives of Erythraeina Welbourn, 1991 and Trombidiina Welbourn, 1991 in Burmese amber. *Cretac. Res.* 89:60–74
74. Koulianos S, Schwarz HH. 1999. Reproduction, development and diet of *Parasitellus fucorum* (Mesostigmata: Parasitidae), a mite associated with bumblebees (Hymenoptera: Apidae). *J. Zool.* 248:267–69
75. Krantz GW. 1998. Reflections on the biology, morphology and ecology of the Macrochelidae. *Exp. Appl. Acarol.* 10:231–58
76. Krantz GW, Poinar GO Jr. 2004. Mites, nematodes and the multimillion dollar weevil. *J. Nat. Hist.* 38:135–41
77. Lachaud JP, Klompen H, Pérez-Lachaud G. 2016. *Macrodirhynchus* mites as parasitoids of invasive ants: an overlooked parasitic association. *Sci. Rep.* 6:29995
78. Lan Q, Lu Z, Ke B, Liao J, Fan QH. 2017. Temperature and humidity effects on physogastric development and reproduction of the mushroom mite *Dolichocybe perniosa* (Acari: Dolichocybidae). *Syst. Appl. Acarol.* 22:1843–48
79. Lara C, Ornelas JF. 2001. Nectar “theft” by hummingbird flower mites and its consequences for seed set in *Moussonia deppeana*. *Funct. Ecol.* 15:78–84
80. Lara C, Ornelas JF. 2002. Effects of nectar theft by flower mites on hummingbird behavior and the reproductive success of their host plant, *Moussonia deppeana* (Gesneriaceae). *Oikos* 96:470–80
81. Lebedeva NV, Lebedev VD. 2007. Diversity of oribatid mites (Acari, Oribatei) and other soil microarthropods in plumage of raptors. *Cauc. Entomol. Bull.* 3:9–18
82. Lesne P. 1986. Mœurs du *Limosina sacra* Meig. [Dipt.]. Phénomènes de transport mutuel chez les animaux articulés. Origine du parasitisme chez les Insectes diptères. *Bull. Soc. Entomol. Fr.* 1:162–65
83. Lindquist EE. 1969. Review of Holarctic tarsonemid mites (Acarina: Prostigmata) parasitizing eggs of ipine bark beetles. *Mem. Entomol. Soc. Can.* 101:S60
84. Lindquist EE. 1986. The world genera of Tarsonemidae (Acari: Heterostigmata): a morphological, phylogenetic, and systematic revision, with a reclassification of family group taxa in the Heterostigmata. *Mem. Entomol. Soc. Can.* 118:S136
85. Lindquist EE, Krantz GW, Walter DE. 2009. Order Mesostigmata. In *A Manual of Acarology*, ed. GE Krantz, DE Walter, pp. 124–232. Lubbock, TX: Texas Tech Univ. Press
86. Lindquist EE, O'Connor BM, Shaw MD, Sidorchuk EA. 2020. Review of the genera *Berlesia* Canestrini, 1884, and *Katidiseius* Fain & Lukoschus, 1983, the subfamily Katydiseiinae Fain & Lukoschus, 1983, and their family group relationships (Acari: Mesostigmata: Gamasina), with description of three new species parasitic on gryllacridid crickets. *Zootaxa* 4857:5–70
87. Liu S, Li J, Guo K, Qiao H, Xu R, et al. 2016. Seasonal phoresy as an overwintering strategy of a phytophagous mite. *Sci. Rep.* 6:25483
88. Lombardero MJ, Klepzig KD, Moser JC, Ayres MP. 2000. Biology, demography and community interactions of *Tarsonemus* (Acarina: Tarsonemidae) mites phoretic on *Dendroctonus frontalis* (Coleoptera: Scolytidae). *Agric. For. Entomol.* 2:193–202
89. Luong LT, Penoni LR, Horn CJ, Polak M. 2015. Physical and physiological costs of ectoparasitic mites on host flight endurance. *Ecol. Entomol.* 40:518–24
90. Luong LT, Subasinghe D. 2016. A facultative ectoparasite attains higher reproductive success as a parasite than its free-living conspecifics. *Exp. Appl. Acarol.* 71:63–70
91. Mackensen J, Bauhus J, Webber E. 2003. Decomposition rates of coarse woody debris—a review with particular emphasis on Australian tree species. *Aust. J. Bot.* 51:27–37
92. Magowski WL. 1994. Discovery of the first representative of the mite subcohort Heterostigmata (Arachnida: Acari) in the Mesozoic Siberian amber. *Acarologia* 35:229–41
93. Magowski WL. 1995. Fossil heterostigmatid mites in amber—85 million year-old arthropod mite relationships. In *The Acari: Physiological and Ecological Aspects of Acari-Host Relationships*, ed. D Kropczyńska, J Boczek, A Tomczyk, pp. 53–58. Krynica, Pol.: DABOR Publ. House

94. McKinney MI, Park YL. 2013. Distribution of *Chaetodactylus krombeini* (Acari: Chaetodactylidae) with *Osmia corniformis* (Hymenoptera: Megachilidae) nests: implications for population management. *Exp. Appl. Acarol.* 60:153–61
95. Moraza ML, Lindquist EE. 2015. Systematics and biology of mites associated with Neotropical hispine beetles in unfurled leaves of *Heliconia*, with descriptions of two new genera of the family Melicharidae (Acari: Mesostigmata: Gamasina: Ascoidea). *Zootaxa* 3931:301–51
96. Moser JC. 1975. Mite predators of the southern pine beetle. *Ann. Entomol. Soc. Am.* 68:1113–16
97. Moser JC, Cross EA. 1975. Phoretomorph: a new phoretic phase unique to the Pyemotidae (Acarina: Tarsonemoidea). *Ann. Entomol. Soc. Am.* 68:820–22
98. Moser JC, Kielczewski B, Wiśniewski J, Bałazy S. 1978. Evaluating *Pyemotes dryas* (Vitzthum 1923) (Acari: Pyemotidae) as a parasite of the southern pine beetle. *Int. J. Acarol.* 4:67–70
99. Nehring V, Teubner H, König S. 2019. Dose-independent virulence in phoretic mites that parasitize burying beetles. *Int. J. Parasitol.* 49:759–67
100. Norton RA. 1980. Observations on phoresy by oribatid mites (Acari: Oribatei). *Int. J. Acarol.* 6:121–30
101. Norton RA. 1994. Evolutionary aspects of oribatid mite life histories and consequences for the origin of the Astigmata. See Ref. 50, pp. 99–135
102. Norton RA, Kethley JB, Johnston DE, O'Connor BM. 1993. Phylogenetic perspectives on genetic systems and reproductive modes of mites. In *Evolution and Diversity of Sex Ratio in Insects and Mites*, ed. D Wrensch, MA Ebbert, pp. 8–99. New York: Chapman & Hall
103. O'Connor BM. 1982. Evolutionary ecology of astigmatid mites. *Annu. Rev. Entomol.* 27:385–409
104. O'Connor BM. 1994. Life-history modifications in astigmatid mites. See Ref. 50, pp. 136–59
105. O'Connor BM. 2009. Cohort Astigmata. In *A Manual of Acarology*, ed. GE Krantz, DE Walter, pp. 565–657. Lubbock, TX: Texas Tech Univ. Press
106. Okabe K. 1999. Morphology and ecology in deutonymphs of non-psoroptid Astigmata. *J. Acarol. Soc. Jpn.* 8:89–101
107. Okabe K. 2013. Ecological characteristics of insects that affect symbiotic relationships with mites. *Entomol. Sci.* 16:363–78
108. Okabe K, Makino S. 2008. Life cycle and sexual mode adaptations of the parasitic mite *Ensliniella parasitica* (Acari: Winterschmidtidae) to its host, the eumenine wasp *Allodynerus delphinalis* (Hymenoptera: Vespidae). *Can. J. Zool.* 86:470–78
109. Okabe K, Makino S. 2008. Parasitic mites as part-time bodyguards of a host wasp. *Proc. R. Soc. B* 275:2293–97
110. Okabe K, Makino S. 2010. Conditional mutualism between *Allodynerus delphinalis* (Hymenoptera: Vespidae) and *Ensliniella parasitica* (Astigmata: Winterschmidtidae) may determine maximum parasitic mite infestation. *Environ. Entomol.* 39:424–29
111. Ontano AZ, Gainett G, Aharon S, Ballesteros JA, Benavides LR, et al. 2020. Taxonomic sampling and rare genomic changes overcome long-branch attraction in the phylogenetic placement of pseudoscorpions. *Mol. Biol. Evol.* 38:2446–67
112. Palevsky E, Soroker V, Weintraub P, Mansour F, Abo-Moch F, Gerson U. 2001. How species-specific is the phoretic relationship between the broad mite, *Polyphagotarsonemus latus* (Acari: Tarsonemidae), and its insect hosts? *Exp. Appl. Acarol.* 25:217–24
113. Pfammatter JA, Coyle DR, Gandhi KJK, Hernandez N, Hofstetter RW, et al. 2016. Structure of phoretic mite assemblages across subcortical beetle species at a regional scale. *Environ. Entomol.* 45:53–65
114. Polak M. 1996. Ectoparasitic effects on host survival and reproduction: the *Drosophila-Macrocheles* association. *Ecology* 77:1379–89
115. Polak M. 1998. Effects of ectoparasitism on host condition in the *Drosophila-Macrocheles* system. *Ecology* 79:1807–17
116. Qu SX, Li HP, Ma L, Song JD, Hou LJ, Lin JS. 2015. Temperature-dependent development and reproductive traits of *Tyrophagus putrescentiae* (Sarcoptiformes: Acaridae) reared on different edible mushrooms. *Environ. Entomol.* 44:392–99
117. Radovsky FJ. 1994. The evolution of parasitism and the distribution of some dermanyssoid mites (Mesostigmata) on vertebrate hosts. See Ref. 50, pp. 186–217

118. Rahmatzadei B, Hajiqanbar H, Mortazavi A, Husemann M. 2021. Global distribution and host range of the endoparasitic mite genus *Locustacarus* (Acari: Podapolipidae) with description of a new species from Iran parasitizing grasshoppers (Orthoptera: Acrididae). *Syst. Parasitol.* 98:487–501
119. Rath W, Delfinado-Baker M, Drescher W. 1991. Observations on the mating behaviour, sex ratio, phoresy and dispersal of *Tropilaelaps clareae* (Acari: Laelapidae). *Int. J. Acarol.* 17:201–8
120. Rettenmeyer CW, Rettenmeyer ME, Joseph J, Berghoff SM. 2011. The largest animal association centered on one species: the army ant *Eciton burchellii* and its more than 300 associates. *Insect Soc.* 58:281–92
121. Robin N, Béthoux O, Sidorchuk E, Cui Y, Li Y, et al. 2016. A Carboniferous mite on an insect reveals the antiquity of an inconspicuous interaction. *Curr. Biol.* 26:1376–82
122. Schäffer S, Kobl Müller S. 2020. Unexpected diversity in the host-generalist oribatid mite *Paraleius leontonychus* (Oribatida, Scheloribatidae) phoretic on Palearctic bark beetles. *PeerJ* 8:e9710
123. Schatz H, Behan-Pelletier VM, O'Connor BM, Norton RA. 2011. Suborder Oribatida van der Hammen, 1968. *Zootaxa* 3148:141–48
124. Schedwill P, Paschkewitz S, Teubner H, Steinmetz N, Nehring V. 2020. From the host's point of view: effects of variation in burying beetle brood care and brood size on the interaction with parasitic mites. *PLOS ONE* 15:e0228047
125. Schmidt AR, Jancke S, Lindquist EE, Ragazzi E, Roghi G, et al. 2012. Arthropods in amber from the Triassic Period. *PNAS* 108:14796–801
126. Schwarz HH, Walz MG. 1996. Pairing, oviposition and development in two sibling species of phoretic mites (Acari: Mesostigmata: Parasitidae: *Poecilochirus* spp.) associated with burying beetles (Coleoptera: Silphidae: *Nicrophorus* spp.). *J. Nat. Hist.* 30:1337–48
127. Seeman OD. 1996. Flower mites and phoresy: the biology of *Hattena panopla* Domrow and *Hattena cometis* Domrow (Acari: Mesostigmata: Ameroseiidae). *Aust. J. Zool.* 44:193–202
128. Seeman OD. 2000. Life cycle, development, feeding and immature life stages of the Fedrizziidae (Mesostigmata: Fedrizzioidea). *Acarologia* 41:39–52
129. Seeman OD. 2008. Systematics and phylogeny of *Cbrysomelobia* species (Acari: Podapolipidae), sexually transmitted parasites of chrysomelid beetles. *Invertebr. Syst.* 22:55–84
130. Seeman OD. 2017. *Megisthanus leviathanicus* sp. nov. (Parasitiformes: Megisthanidae), the largest known Mesostigmata, a symbiont of the beetle *Mastachilus australasicus* (Coleoptera: Passalidae). *Int. J. Acarol.* 43:263–85
131. Seeman OD. 2021. Contrasting species diversification of *Eutarsopolipus* (Acariformes: Podapolipidae) on *Castelnaudia* and *Notonomus* (Coleoptera: Carabidae). *Zootaxa* 4971:174
132. Seguy E. 1950. Phoresie et rôle pathogène chez les insectes diptères. *Eos* 1950:315–24
133. Shaw M. 2010. Post-larval stages of *Ascoschoengastia* (*Laurentella*) *lorius* (Gunther) (Acariformes: Trombiculidae) provide evidence for a nest-based life history. *Zootaxa* 2680:55–64
134. Sidorchuk EA. 2018. Mites as fossils: forever small? *Int. J. Acarol.* 44:349–59
135. Smith BP. 1998. Loss of larval parasitism in parasitengonine mites. *Exp. Appl. Acarol.* 22:187–99
136. Smith IM, Oliver DR. 1986. Review of parasitic associations of larval water mites (Acari: Parasitengona: Hydrachnida) with insect hosts. *Can. Entomol.* 118:407–72
137. Sun SJ, Kilner RM. 2020. Temperature stress induces mites to help their carrion beetle hosts by eliminating rival blowflies. *eLife* 9:e55649
138. Theron-De Bruin N, Dreyer LL, Ueckermann EA, Wingfield MJ, Roets F. 2018. Birds mediate a fungus-mite mutualism. *Microb. Ecol.* 75:863–74
139. Trach VA. 2013. On the morphology, biology, and distribution of *Lobogynioides andreinii* (Acari, Mesostigmata, Diplogyniidae). *Entomol. Rev.* 93:105–12
140. Treat AE. 1975. *Mites of Moths and Butterflies*. Ithaca, NY: Comstock
141. Walter DE, Lindquist EE. 1995. The distributions of parthenogenetic ascid mites (Acari: Parasitiformes) do not support the biotic uncertainty hypothesis. *Exp. Appl. Acarol.* 19:434–42
142. Walter DE, Proctor HC. 2013. *Mites: Ecology, Evolution and Behaviour*. Berlin: Springer. 2nd ed.
143. Walter DE, Seeman OD. 2017. A new species of *Paracarophenax* (Acariformes: Acarophenacidae) with a new means of phoretic attachment. *Int. J. Acarol.* 43:329–35

144. Wheeler WM. 1919. The phoresy of *Antherophagus*. *Psyche* 26:145–52
145. Wilson DS, Knollenberg WG. 1987. Adaptive indirect effects: the fitness of burying beetles with and without their phoretic mites. *Evol. Ecol.* 1:139–59
146. Wohltmann A. 2001. The evolution of life histories in Parasitengona (Acari: Prostigmata). *Acarologia* 41:145–204
147. Yoder JA, Jajack AJ, Tomko PM, Rosselot AE, Gribbins KM, Benoit JB. 2012. Pollen feeding in *Balaustium murorum* (Acari: Erythraeidae): visualization and behaviour. *Int. J. Acarol.* 38:641–47
148. Zhang ZQ, Fan QH, Pesic V, Smit H, Bochkov AV, et al. 2011. Order Trombidiformes Reuter, 1909. *Zootaxa* 3148:129–38
149. Zhang ZQ, Gerson U. 1995. *Eustigmaeus johnstoni*, new species (Acari: Stigmaeidae), parasitic on phlebotomine sandflies (Diptera: Psychodidae). *Tijdschr. Entomol.* 138:297–301